RESEARCH ARTICLE

WASP-Arp2/3-dependent actin polymerization influences fusogen localization during cell-cell fusion in Caenorhabditis elegans embryos

Yan Zhang*, Yihong Yang*, Zhiwen Zhu and Guangshuo Ou†

ABSTRACT

Cell-cell fusion is essential for development and physiology. Actin polymerization was implicated in the Caenorhabditis elegans fusogen EFF-1 engagement in a reconstituted Drosophila cell culture system, and the actin-binding protein spectraplakin links EFF-1 to the actin cytoskeleton and promotes cell-cell fusions in C. elegans larvae. However, it remains unclear whether and how fusogens and the actin cytoskeleton are coordinated in C. elegans embryos. Here, we used live imaging analysis of GFP knock-in and RNAi embryos to study the embryonic cell-cell fusions in C. elegans. Our results show that the inhibition of WASP-Arp2/3-dependent actin polymerization delays cell-cell fusions. EFF-1 is primarily distributed in intracellular vesicles in embryonic fusing cells, and we find that the perturbation of actin polymerization reduces the number of EFF-1-positive vesicles. Thus, the actin cytoskeleton differentially promotes cell-cell fusion by regulating fusogen localization to the fusing plasma membrane in larvae or to intracellular vesicles in embryos.

KEY WORDS: Actin polymerization, Arp2/3, Fusogen, Cell-cell fusion, Caenorhabditis elegans

INTRODUCTION

Cell-cell fusion is involved in various developmental and physiological events, ranging from sexual reproduction, myogenesis, bone remodeling and immune responses (Chen, 2011; Chen and Olson, 2005; Podbilewicz, 2014). The Caenorhabditis elegans fusogenic protein EFF-1 was isolated from genetic screens of epithelial fusion failure and the eff-1 gene encodes a type I single transmembrane protein (Mohler et al., 2002; Podbilewicz, 2006; Sapir et al., 2007). EFF-1 shares structural homology with the green alga Chlamydomonas reinhardtii HAPLESS 2/GENERATIVE CELL SPECIFIC 1 (HAP2/GCS1) family of proteins and viral class II fusion proteins (Fédry et al., 2006; Sens et al., 2010; Podbilewicz, 2014). The largest epithelial syncytium contains 139 nuclei, which is formed when embryonic cell-cell fusion merges 23 cells and the remaining 116 cells fuse in larvae (Podbilewicz and White, 1994; Sulston and Horvitz, 1977). To understand whether the actin cytoskeleton plays a general role in C. elegans cell-cell fusion, we examined the dorsal hyp7 cell fusion in Arp2/3 and WASP RNAi embryos. The previous RNAi of the ARX-2 subunit in the Arp2/3 complex did not perturb hyp7 cell fusion, and the animals could survive for >3 days (Patel et al., 2008). Our RNAi treatment caused 100% animal lethality within 2 days (n=200 for each genotype of WASP, Arp2/3 or WAVE), which is suggestive of a more potent depletion. To visualize the cell boundary of two fusing epithelial cells, we constructed a knock-in animal of

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.

Received 14 May 2017; Accepted 12 July 2017


Tsinghua-Peking Center for Life Sciences, School of Life Sciences and MOE Key Laboratory for Protein Science, Tsinghua University, Beijing 100084, China.
*These authors contributed equally to this work
†G.O., 0000-0003-1512-7824

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.
Fig. 1. See next page for legend.
TagRFP-tagged DLG-1 that is orthologous to the Drosophila Discs large and localizes at apical adherens junctions in all epithelia (Fig. 1B) (Smurova and Podbilewicz, 2016). We also used GFP or TagRFP-tagged small GTPase MIG-2 to visualize the plasma membrane (Fig. 2D and Fig. 3A) (Ou and Vale, 2009). We performed fluorescence time-lapse imaging analysis of embryonic cell-cell fusions. A total of 17 dorsal hyp7 precursor cells merged their plasma membranes to form the hyp7 syncytium (Fig. 1A). The loss of DLG-1::TagRFP fluorescence at the border of two hyp7 cells indicates the completion of cell-cell fusion (Fig. 1B).

In DLG-1::TagRFP embryos, the red fluorescent cell boundary between two fusing hyp7 cells progressively disappeared (Fig. 1B). At the twofold developmental stage, only 4±1 (n=12) borders could be detected, indicative of the completion of 13±1 hyp7 cell-cell fusions. However, all the DLG-1::TagRFP fluorescence was retained in eff-1(ok1021) mutant embryos at the same development stage (Fig. 1B). RNAi treatment of arx-2, arx-3 or wsp-1 delayed 9±2, 8±2 or 9±1 hyp7 precursor cell fusions at the twofold stage, respectively (n=10-12; Fig. 1B-D; Movie 1). The delayed cell-cell fusions in WASP and Arp2/3 RNAi embryos may result from an incomplete depletion of WASP or Arp2/3 using RNAi. Importantly, 11±3 and 11±2 hyp7 precursor cells (n=10-12) did not fuse in arx-2; arx-3 and arx-2; wsp-1 double RNAi embryos at this stage, indicating an enhancement of cell fusion defects (Fig. 1B-D). Consistent with this notion, the larval cell-cell fusion defects were also enhanced in wsp-1 and arx-2 double conditional knockouts, presumably because of the additive disruption of WASP and Arp2/3 (Yang et al., 2017). In agreement with the previous data, RNAi of wee-1 did not cause any apparent defects of cell-cell fusion in embryos (Fig. 1B-D), which indicates that WASP, but not WAVE, is the essential actin nucleation promoting factor during cell-cell fusions. Thus, the WASP-Arp2/3-mediated actin polymerization is involved in cell-cell fusion in C. elegans embryos.

The actin cytoskeleton regulates the vesicular localization of EFF-1 in embryos

To further investigate the coordination between EFF-1 and actin polymerization in embryos, we generated EFF-1::GFP and ARX-2::TagRFP double knock-in animals. We performed studies by quantifying the percentage of endogenous EFF-1::GFP puncta that were associated with ARX-2::TagRFP puncta in hyp7 precursor cells (Fig. 2A-C; Movie 2). Forty-five percent of EFF-1 puncta overlapped with ARX-2 puncta (Fig. 2C). Moreover, our kymograph analysis revealed that EFF-1::GFP and ARX-2::TagRFP move together in some migratory puncta (Fig. 2B). To examine whether actin polymerization is required for EFF-1::GFP distribution in hyp7 precursor cells, we compared the EFF-1::GFP puncta density in WT or arx-2 and wsp-1 RNAi embryos. WT embryos contained 0.17±0.04 EFF-1::GFP puncta per µm², whereas RNAi of arx-2, wsp-1 or both reduced the puncta density to 0.05±0.02, 0.04±0.01 and 0.02±0.01 per µm², respectively (Fig. 3A,B). When we quantified the overall levels of EFF-1::GFP, we did not observe any statistically significant changes in the fusing cells of arx-2 or wsp-1 RNAi embryos (Fig. 3C). These results indicate that WASP and Arp2/3 do not affect the expression or the
stability of EFF-1, but regulate EFF-1 vesicular localization, suggesting that EFF-1 may be evenly distributed to the cytoplasm in C. elegans arx-2 or wsp-1 embryos.

Considering that EFF-1::GFP forms puncta in the postembryonic seam V.a (anterior daughter cell of the V cell) cells, we examined whether WASP is also involved in the membrane trafficking of EFF-1 in larval seam cells (Yang et al., 2017). Using the same protocol as for EFF-1 quantification in embryos, we found that the WT V.a cells and wsp-1-deficient cells contained 0.14±0.03 and 0.12±0.02 EFF-1::GFP puncta per μm², respectively (mean±s.d., n=16, no statistical significance based on Student’s t-test). Although the transmembrane protein EFF-1 must be synthesized in the endoplasmic reticulum and undergo membrane trafficking to arrive on the plasma membrane in both embryonic and larval cells, our results indicate that the underlying regulatory mechanisms appear to be different at distinct developmental stages (Fig. 3D) (Yang et al., 2017).

EFF-1 has distinct localization patterns at different developmental stages in forming a single epithelial syncytium. Although the actin cytoskeleton is generally involved in this process, it facilitates EFF-1 localization in a distinct manner in embryos and larvae. These results indicate that cell-cell fusion may be more complex than is currently appreciated. A previous study revealed that EFF-1 is removed from the plasma membrane by RAB-5 and dynamin-mediated endocytosis in embryos (Smurova and Podbilewicz, 2016). We showed that ARX-1 puncta always localized at the borders of the fusing hyp7 cells (Fig. 2D, n>50 for each marker; Movie 3) and that EFF-1 puncta could occasionally be detected at the border (Yang et al., 2017). Because WASP and Arp2/3 are essential for cell-cell fusion (Fint-Karalar and Welch, 2011), the reduction of EFF-1 puncta in the absence of WASP or Arp2/3 can be explained by defective endocytosis. However, the perturbation of endocytosis led to the hyperfusion phenotype (Smurova and Podbilewicz, 2016) rather than the delay of cell-cell fusion observed in WASP or Arp2/3 RNAi embryos. Given that Arp2/3-based actin nucleation is also required for ER-to-Golgi transport (Campellone et al., 2008), and that EFF-1 puncta colocalize with the Golgi (Smurova and Podbilewicz, 2016), Arp2/3-dependent secretory sorting may facilitate the trafficking of EFF-1 from the ER to the plasma membrane, thereby promoting cell-cell fusion.

**MATERIALS AND METHODS**

**C. elegans strains, genetics and DNA manipulations**

Strains were maintained on nematode growth medium (NGM) plates seeded with *Escherichia coli* OP50 following standard protocols at 20°C. CRISPR-Cas9-assisted knock-in animals were generated and analyzed as described previously (Dickinson et al., 2013; Shen et al., 2014). To construct knock-in repair template plasmids, we amplified the 1-1.5 kb upstream and downstream homologous arms from the N2 genomic DNA and inserted them into *pPD95.77* with an In-Fusion Advantage PCR Cloning Kit (Clontech, Mountain View, USA). To avoid the cleavage of the homologous repair template by Cas9, synonymous mutations were introduced to the Cas9 target site of the template. The sgRNA plasmid and the knock-in repair template plasmids, we amplified the 1-1.5 kb upstream and downstream homologous arms from the N2 genomic DNA and inserted them into *pPD95.77* with an In-Fusion Advantage PCR Cloning Kit (Clontech, Mountain View, USA). To avoid the cleavage of the homologous repair template by Cas9, synonymous mutations were introduced to the Cas9 target site of the template. The sgRNA plasmid and the knock-in repair template plasmids were co-injected into N2 animals. The knock-in worms were selected and examined by PCR and Sanger sequencing. Transgenic knock-in template plasmid were co-injected into N2 animals. The knock-in worms were selected and examined by PCR and Sanger sequencing. Transgenic strains are listed in Tables S1-S4.

**Feeding RNAi**

Feeding RNAi was performed as described previously (Kamath et al., 2001). For arx-2, arx-3, arx-5, arx-6, arx-7 and wsp-1, the clones from the


