Cardiac-enriched BAF chromatin-remodeling complex subunit Baf60c regulates gene expression programs essential for heart development and function

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ABSTRACT

How chromatin-remodeling complexes modulate gene networks to control organ-specific properties is not well understood. For example, Baf60c (Smard3) encodes a cardiac-enriched subunit of the SWI/SNF-like BAF chromatin complex, but its role in heart development is not fully understood. We found that constitutive loss of Baf60c leads to embryonic cardiac hypoplasia and pronounced cardiac dysfunction. Conditional deletion of Baf60c in cardiomyocytes resulted in postnatal dilated cardiomyopathy with impaired contractile function. Baf60c regulates a gene expression program that includes genes encoding contractile proteins, modulators of sarcomere function, and cardiac metabolic genes. Many of the genes deregulated in Baf60c null embryos are targets of the MEF2/SRF co-factor Myocardin (MYOCID). In a yeast two-hybrid screen, we identified MYOCID as a BAF60c interacting factor; we showed that BAF60c and MYOCID directly and functionally interact. We conclude that Baf60c is essential for coordinating a program of gene expression that regulates the fundamental functional properties of cardiomyocytes.

KEY WORDS: Chromatin remodeling, Embryo, Gene regulation, Heart

INTRODUCTION

Transcription factor networks control cardiac morphogenesis and cell specification (Bruneau, 2013; Evans et al., 2010), including the coordinated regulation of genes encoding the proteins involved in sarcomere function (Creemers et al., 2006; Niu et al., 2008). While undergoing complex morphogenetic changes, the developing heart supports embryonic circulation. The contractile function of the heart adapts quickly to the dramatic changes in circulation that occur after birth and, subsequently, must adapt to fluctuating physiology and stress. The transcriptional regulation of cardiac gene expression continues during postnatal heart growth and cardiomyocyte maintenance (Huang et al., 2009; Oka et al., 2006).

Chromatin-remodeling complexes are critical regulators of cardiac gene expression, in many cases modulating the activity of DNA-binding transcription factors (Chang and Bruneau, 2012). For example, histone deacetylases (HDACs) and bromodomain-containing factors have important roles in cardiac gene regulation and remodeling, and have been proposed as potential drug targets (Anand et al., 2013; McKinsey, 2012). BRG1/BRM-associated factor (BAF) complexes are ATP-dependent chromatin remodeling complexes related to the yeast SWI/SNF complex, and are indispensable for mammalian development (Hota and Bruneau, 2016). BAF complexes orchestrate many aspects of heart development, and genetically interact with cardiac transcription factors to finely modulate cardiac gene expression (Hang et al., 2010; Takeuchi et al., 2011). Combinatorial assembly of different polymorphic subunits can generate hundreds of potential BAF complexes, and offer precise control of developmental processes (Chang and Bruneau, 2012; Ho and Crabtree, 2010). BAF60c (also known as SMARCD3) is a polymorphic subunit of the BAF complex, which is expressed preferentially in the developing heart (Lickert et al., 2004). In vivo RNAi knockdown in mouse embryos suggested that Baf60c is essential for embryonic heart development (Lickert et al., 2004), and together with the cardiac transcription factors TBX5, NKX2-5 and GATA4, BAF60c can induce non-cardiac mesoderm to differentiate into cardiomyocytes (Lou et al., 2011; Takeuchi and Bruneau, 2009).

In this study, we examined the role of Baf60c in embryonic and postnatal heart development in a Baf60c conditional knockout mouse line. We showed that Baf60c is essential for cardiac growth and cardiomyocyte function at several stages of embryonic development, by regulating broad networks of genes encoding proteins essential for function of the contractile apparatus. Many of the dysregulated genes are targets of the MEF2 co-factor MYOCID, and we identified MYOCID as a BAF60c-interacting protein. Our work shows that Baf60c is an important modulator of the fundamental program of gene expression essential for cardiac structure and function.

RESULTS

Construction of Baf60c conditional knockout mouse line
Baf60c is expressed at embryonic day (E) 7.5 in the early cardiac precursors of the cardiac crescent, and its expression is maintained...
throughout development in the myocardium (Lickert et al., 2004). To understand the function of Baf60c at different developmental stages, we developed a conditional allele of Baf60c in the mouse. A targeting construct with a pair of loxP sites flanking exon 1–4 was introduced into embryonic stem (ES) cells (Fig. 1A). Transgenic mice generated from the targeted ES cells (Baf60clox/lox) had normal phenotypes and lifespans and, thus, were treated as wild type. By crossing them with pCAGGS-Cre mice, which constitutively express Cre recombinase, exons 1–4 of Baf60c were deleted to generate Baf60c+/- mice (Fig. 1A). No obvious defects were observed in Baf60c+/- mice. Homozygous null Baf60c−/− embryos were recovered at E9.5 (Fig. 1B), and by whole mount in situ hybridization, no Baf60c mRNA was detectable in Baf60c−/− embryos (Fig. 1C).

**Baf60c deletion results in a hypoplastic heart and embryonic demise**

Baf60c−/− embryos were recovered alive and with roughly normal morphology at different stages of timed pregnancies until E12.5–E14.5. At E14.5, most Baf60c−/− embryos were dead, with broad regions of hemorrhage. Backcrossing into C57Bl/6 for 10 generations led to a more consistent phenotype, with survival only until E12.5–13.5. To determine the cause of embryonic death and to identify potential cardiac phenotypes, Baf60c−/− embryos were harvested for histological analysis. Optical projection tomography showed that mixed background E12.5 Baf60c−/− embryonic hearts had dilated inner chambers and underdeveloped interventricular septa (Fig. 2A). At E11.5, Baf60c−/− C57Bl/6 embryonic hearts had a more severe and penetrant phenotype, with a thin compact layer and fewer or less well-developed trabeculae (Fig. 2B), impaired atrioventricular cushion formation, and reduced atrial septum growth. In the few surviving E14.5 Baf60c−/− mixed background embryos, ventricular free walls were much thinner than wild type (Fig. 2C), and the interventricular septum was disorganized, leading to ventricular septal defects. Based on the intrinsic cardiac phenotypes, we conjectured that circulatory failure and hemorrhage were the result of impaired cardiac function of Baf60c−/− embryos.

To identify the possible cause of cardiac hypoplasia in Baf60c knockouts, proliferation of cardiomyocytes was assessed by staining with Ki67 antibody. Immunostaining detected fewer Ki67+ cardiomyocytes in E12.5 Baf60c−/− ventricles than in

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**Fig. 1. Construction of Baf60c knockout mouse line.** (A) Schematic representation of targeting DNA introduced into wild-type (WT) Baf60c locus. Correctly targeted ES cells were identified with probe located outside of the homologous arm. Cre-mediated excision removed exons 1–4 and resulted in Baf60c+/- . (B) Left: Southern blot of digested ES cell DNA with an external probe outside of the targeting DNA. WT and targeted band size are as described in A. Right: genotype PCR showing the band size differences of WT, Baf60c+/- (KO) and heterozygous Baf60c+/- . (C) Whole-mount in situ hybridization using full-length Baf60c probe detected no signals in genotyped homozygous Baf60c−/− embryos (n>3), indicating complete deletion. E, EcoRI; fb, forebrain; ht, heart; sm, somites.
wild type (Fig. 2D). Quantitation confirmed that, in wild-type (WT) hearts, there were 32±9% Ki67+ ventricular cardiomyocytes, and in Baf60c−/− hearts, 25±5% were positive (n=4; P<0.05 by unpaired two-tailed t-test). There was no evidence of increased apoptosis.

The embryonic heart begins to pump blood from the linear heart tube stage onwards, and its contractile function is essential for fetal life. To determine if cardiac function was affected by Baf60c deletion, we used high frequency ultrasound echocardiography (Zhou et al., 2002) to evaluate contractile parameters of E13.5 mixed background embryos in utero (Table 1). No regurgitation between atria and ventricles was observed in Baf60c−/− embryos, which indicates that cardiac valves had formed and were fully functional. However, the left ventricle fraction shortening (LVFS) of Baf60c−/− hearts was reduced, suggesting impaired systolic function. The inter-ventricular septal fractional thickening (IVSFT) was lower than in the hearts of WT and Baf60c+/− embryos, indicating reduced myocardial contraction. The E:A ratios of Baf60c−/− hearts for both the left and right ventricles were also significantly increased. This may indicate impaired cardiac relaxation (Zhou et al., 2003). Overall, echocardiography showed that loss of Baf60c affected the morphology and dimensions of the heart and, concomitantly, its contractile function.

Since Baf60c is expressed at sites outside the developing heart, such as extraembryonic tissues and neural tube, we assessed the tissue specificity of the Baf60c−/− phenotype by crossing Baf60cflox/flox mice with Nkx2-5::Cre mice. Nkx2-5::Cre deletes loxP-flanked DNA from E8.0 in all cardiac precursors (Moses et al., 2001). No live Baf60cflox−/−;Nkx2-5::Cre embryos were recovered after E14.5. By E14.5 Baf60cflox−/−;Nkx2-5::Cre embryo ventricle free walls and ventricular septum were much thinner than in littermate controls (Fig. 3). The ventricular septum was also poorly organized. This hypoplastic cardiac phenotype of Baf60c−/−; Nkx2-5::Cre embryos phenocopies the least affected E14.5 Baf60c−/− embryos, and suggests that the cardiac defects of the Baf60c knockout is a direct effect of Baf60c absence in the heart.
instead of an indirect effect from Baf60c loss in other tissues like neural tube and somites. Indeed, no other embryonic defects were noted in Baf60c<sup>fl<sup>ox<sup>−</sup>:Nkx2-5::Cre</sup> embryos. The constitutive null phenotype thus likely reflects primary loss of Baf60c in the developing heart, but as it is comparatively more severe might also reflect an earlier function in precursors that do not yet express Nkx2-5 (Devine et al., 2014).

Loss of Baf60c in cardiomyocytes results in postnatal cardiomyopathy

After birth, heart development switches from cell proliferation to hypertrophic growth. The structure and physiological function of the myocardium undergo a series of changes to adapt to a new hemodynamic environment. We deleted Baf60c in the myocardium at later developmental stages by crossing the Baf60c<sup>fl<sup>ox<sup>−</sup> allele with Myh6::Cre (Agah et al., 1997). This manipulation bypassed the embryonic lethality of the constitutive deletion, as Baf60c<sup>−/−</sup>;Myh6::Cre (Baf60c<sup>Myh6KO</sup>) mice were born alive and showed no obvious morphological changes before postnatal day (P)7. After P7, some of the Baf60c<sup>Myh6KO</sup> pups had delayed growth, compared with their littermates, and died before weaning. Other Baf60c<sup>Myh6KO</sup> mice survived after weaning without obvious morphological defects, but, at 4–6 weeks, exhibited symptoms of heart failure, including weight loss, reduced activity level, hunched back and labored breath. The remaining Baf60c<sup>Myh6KO</sup> mice appeared normal, but died suddenly. All Baf60c<sup>Myh6KO</sup> mice died before 4 months of age (Fig. 4A).

To investigate the reason for the early mortality in Baf60c<sup>Myh6KO</sup> mice, their hearts were dissected at different ages for morphology and histology analysis. At all the observed stages (P10, P21 and 8 weeks), the hearts of Baf60c<sup>Myh6KO</sup> mice were larger than controls (Fig. 4B). Histology revealed chamber dilation (Fig. 4B). Masson’s trichrome staining detected broad myocardium interstitial fibrosis in the Baf60c<sup>Myh6KO</sup> myocardium, while this was not observed in any other genotypes (Fig. 4B, lower panels). A high level of apoptosis was also detected in myocardium of adult Baf60c<sup>Myh6KO</sup> mice (Fig. 4C).

The chamber dilation and fibrosis observed in the hearts of Baf60c<sup>Myh6KO</sup> mice raised the question of whether cardiac function was also affected. We measured cardiac contractile function of

Table 1. High frequency echocardiography evaluation of E13.5 embryos

<table>
<thead>
<tr>
<th></th>
<th>WT (n=4)</th>
<th>Baf60c&lt;sup&gt;1/2&lt;/sup&gt;−− (n=5)</th>
<th>Baf60c&lt;sup&gt;−/−&lt;/sup&gt;−− (n=5)</th>
<th>P-value</th>
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<tbody>
<tr>
<td>LV ESD (mm)</td>
<td>0.67±0.03</td>
<td>0.72±0.02</td>
<td>0.79±0.02</td>
<td>P=0.0087</td>
</tr>
<tr>
<td>LV EDD (mm)</td>
<td>0.94±0.04</td>
<td>0.99±0.03</td>
<td>1.03±0.03</td>
<td>P=0.01 (Het versus KO)</td>
</tr>
<tr>
<td>LV FS (%)</td>
<td>28±2</td>
<td>27±1</td>
<td>27±0.02</td>
<td>P=0.002 (WT versus KO)</td>
</tr>
<tr>
<td>RV ESD (mm)</td>
<td>0.64±0.03</td>
<td>0.64±0.04</td>
<td>0.69±0.03</td>
<td>NS</td>
</tr>
<tr>
<td>RV EDD (mm)</td>
<td>0.85±0.02</td>
<td>0.89±0.03</td>
<td>0.92±0.02</td>
<td>NS</td>
</tr>
<tr>
<td>RV FS (%)</td>
<td>25±3</td>
<td>32±2</td>
<td>24±3</td>
<td>NS</td>
</tr>
<tr>
<td>IVSTes (mm)</td>
<td>0.32±0.02</td>
<td>0.36±0.03</td>
<td>0.25±0.03</td>
<td>P=0.0023</td>
</tr>
<tr>
<td>IVSTed (mm)</td>
<td>0.22±0.03</td>
<td>0.25±0.03</td>
<td>0.20±0.02</td>
<td>NS</td>
</tr>
<tr>
<td>IVSFT</td>
<td>40±4</td>
<td>41±4</td>
<td>26±6</td>
<td>P=0.0084 (WT versus KO)</td>
</tr>
<tr>
<td>P&lt;0.006 (Het versus KO)</td>
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<tr>
<td>Mitral peak E/A</td>
<td>0.12±0.04</td>
<td>0.25±0.05</td>
<td>0.32±0.04</td>
<td>P=0.02</td>
</tr>
<tr>
<td>Tricuspid peak E/A</td>
<td>0.19±0.03</td>
<td>0.17±0.03</td>
<td>0.34±0.04</td>
<td>P=0.008</td>
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</tbody>
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IVSFT, inter-ventricular septal fractional thickening; IVSTed, end-diastolic inter-ventricular septum thickness; IVSTes, end-systolic inter-ventricular septum thickness; LV EDD, left ventricular end-diastolic diameter; LV ESD, left ventricular end-systolic diameter; LV FS, left ventricular fractional shortening; RV EDD, right ventricular end-diastolic diameter; RV ESD, right ventricular end-systolic diameter; RV FS, right ventricular fractional shortening. Peak E/A, the ratio of peak velocities of the early diastolic waveform (E wave) to the late diastolic waveform during atrial contraction (A wave) at either mitral or tricuspid orifices. P-value indicates significance assessed by ANOVA.
8-week-old mice that lacked outward signs of heart failure or growth delay, using high frequency echocardiography (Table 2, n=6). Confirming the histological results, the left ventricles of Baf60cMyh6KO mice were prominently dilated, and the anterior and posterior ventricle walls of Baf60cMyh6KO mice were thinner and the chamber contraction ratio decreased. The aortic time-velocity integral (TVI, which measures the distance traveled by a volume of blood during a time interval) increased, probably because of the

Table 2. Echocardiography analysis of the cardiac contractile function Baf60cMyh6KO mice at 8 weeks of age

<table>
<thead>
<tr>
<th></th>
<th>Baf60c+/+</th>
<th>Baf60c+/+; Myh6::Cre</th>
<th>Baf60c−/−</th>
<th>Baf60c−/−; Myh6::Cre</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Awed (mm)*</td>
<td>0.93±0.03</td>
<td>0.88±0.03</td>
<td>0.80±0.05</td>
<td>0.70±0.04*</td>
<td>P=0.0027</td>
</tr>
<tr>
<td>AWes (mm)*</td>
<td>1.27±0.05</td>
<td>1.19±0.05</td>
<td>1.18±0.08</td>
<td>0.96±0.05*</td>
<td>P=0.0082</td>
</tr>
<tr>
<td>PWed (mm)*</td>
<td>0.84±0.04</td>
<td>0.73±0.04</td>
<td>0.76±0.03</td>
<td>0.66±0.04*</td>
<td>P=0.013</td>
</tr>
<tr>
<td>PWes (mm)*</td>
<td>1.10±0.04</td>
<td>1.02±0.07</td>
<td>1.07±0.03</td>
<td>0.90±0.06*</td>
<td>P=0.046</td>
</tr>
<tr>
<td>EDD (mm)*</td>
<td>4.18±0.09</td>
<td>4.19±0.02</td>
<td>4.34±0.08</td>
<td>5.01±0.18*</td>
<td>P&lt;0.001</td>
</tr>
<tr>
<td>ESD (mm)*</td>
<td>3.06±0.13</td>
<td>3.09±0.09</td>
<td>2.99±0.04</td>
<td>3.85±0.25*</td>
<td>P=0.0021</td>
</tr>
<tr>
<td>FS (%)</td>
<td>26±2</td>
<td>31±1</td>
<td>26±2</td>
<td>23±2</td>
<td>P&lt;0.05</td>
</tr>
<tr>
<td>Aortic diameter (mm)</td>
<td>1.17±0.02</td>
<td>1.15±0.02</td>
<td>1.14±0.03</td>
<td>1.11±0.01</td>
<td>NS</td>
</tr>
<tr>
<td>Aortic TVI (mm)*</td>
<td>35.4±1.9</td>
<td>38.8±2.2</td>
<td>37.4±1.1</td>
<td>43.1±1.8*</td>
<td>P=0.043</td>
</tr>
<tr>
<td>CO (ml/min)*</td>
<td>14.8±0.6</td>
<td>15.0±0.9</td>
<td>12.6±0.7</td>
<td>12.4±0.6*</td>
<td>P=0.032</td>
</tr>
</tbody>
</table>

AWed, anterior wall thickness at end diastole; AWes, anterior wall thickness at end-systole; EDD, end-diastolic diameter; ESD, end-systolic diameter; FS, fractional shortening; PWed, posterior wall thickness at end-diastole; PWes, posterior wall thickness at end systole; TVI, time-velocity integral; CO, cardiac output. Data are mean±s.e.m. P-value indicates significance assessed by ANOVA. Asterisks indicate significantly different values for individual groups.
enlarged ventricle volume. The fractional shortening (FS) and cardiac output were reduced, consistent with the cardiac failure symptoms of Baf60cMyh6KO mice. We performed electrocardiogram analysis to measure the conduction function of Baf60cMyh6KO mice (Fig. 4D, Table 3; n=5–6). Compared with other genotypes, Baf60cMyh6KO mice had significantly slower heart rates, shortened conduction time through the atrioventricular (AV) node (PR interval), and prolonged QRS duration, suggesting longer depolarization-repolarization time of the ventricle. P wave height, which indicates atrial depolarization, was reduced. Thus, clear and significant conduction defects accompany contractile deficiency in Baf60cMyh6KO mice.

Myofibrillar defects of Baf60c KO cardiomyocytes
The cardiac structural and functional defects in Baf60c−/− are a reflection of an underlying cellular defect. To address this, we used electron microscopy to observe cardiomyocyte ultrastructure. At E12.5, sarcomeres of Baf60c−/− hearts were disarrayed, and the thick and thin filaments were discontinuous and poorly aligned. Z-disks were loosely packed and did not have clearly defined borders as in WT sarcomeres. The I bands (thick-filament free zone) and the M bands (myosin head free zone of the thick filaments) located in the middle of sarcomere were almost undetectable (Fig. 5A, top panel). Similar defects also existed in adult Baf60c−/− cardiomyocytes. The sarcomere length in adult hearts (the length between two adjacent Z-disks) was significantly shorter in Baf60c−/− hearts than WT mice (Fig. 5B).

We examined the distribution of several important structural proteins in cardiomyocytes by immunofluorescence deconvolution microscopy, and found that the localization of Desmin in Z-disks of embryonic cardiomyocytes was disturbed in Baf60c−/− hearts (Fig. 5C). In adult Baf60c−/− hearts, localization of Desmin in intercalated discs was also reduced (Fig. 5D), and the pattern of Desmin localization was perturbed (poorly aligned). These observations are similar to what was observed by electron microscopy and, together, show disrupted myofibril alignment and sarcomere structure in the absence of Baf60c.

Cardiac gene expression program regulated by Baf60c
To identify genes regulated by Baf60c, we used RNAseq to analyze total RNA prepared from Baf60c−/− hearts and control hearts harvested at E10.5 and E12.5, and from P7 Baf60c−/− and control hearts. We identified 788 genes that were differentially expressed by at least 1.25-fold (P<0.05) in at least one stage versus wild type (Fig. 6A; Supplemental Datasets 1 and 2). Among all the genes and all the analyzed stages, 132 genes were upregulated, and 175 were downregulated in embryonic and adult Baf60c−/− hearts (Fig. 6D). A broader range of cardiac function-related genes were deregulated in the Baf60c−/− hearts, including Gja3, Myl1, Myl4, Myl7, and Tnn1. The postnatal deletion of Baf60c also resulted in induction of Nppa, as might be expected in a cardiomyopathic heart (Houweling et al., 2005), but the induction was mild (only twofold increase), indicating a potential deficiency in upregulation of this marker of cardiac stress. In fact, the usual cardiac stress-responsive genes were not present in the Baf60c−/− cardiac gene expression program. Gene ontology (GO) analysis of genes repressed by Baf60c in postnatal heart enriched for biological processes involved in broad developmental processes and extracellular structure organization (Fig. 6C). However, Baf60c-activated genes were enriched for muscle system processes, regulation of muscle cell differentiation, muscle contraction, and sarcomere and actin cytoskeleton organization (Fig. 6D). An enrichment of cell-cycle-related genes was also apparent; it is not clear what this signifies and may reflect a role for Baf60c in regulating perinatal proliferation, which was not addressed in this study. These results collectively suggest that Baf60c is required for proper expression of genes encoding components or regulators of the contractile apparatus.

The analysis of gene expression in whole hearts has the disadvantage that a heterogeneous mix of cells may prevent clear identification of the full set of Baf60c-regulated genes, and also that some changes in gene expression may be secondary to altered hemodynamics. We compared the set of genes altered in the Baf60c mutant hearts with RNAseq analysis of cardiac precursors and cardiomyocytes differentiated in vitro from WT and Baf60c−/− ES cells (Hota et al., 2017 preprint). Considerable overlap was found for the E12.5 KO hearts and ES cell-derived cardiac precursors differentially expressed genes, and more significant overlap was found for E12.5 KO and Baf60c−/− hearts with ES cell-derived cardiomyocyte (Fig. 6E). These comparisons show that both in vitro and in vivo, Baf60c regulates a set of genes important for cardiac morphogenesis and function.

BAF60c functionally interacts with Myocardin
We previously identified TBX5 and NKX2.5 as potential BAF60c-interacting proteins (Lickert et al., 2004). Here we used GST pulldown to show that these interactions can be direct (Fig. 7A). We mapped the BAF60c interaction domain to an N-terminal region that contains a nuclear localization signal sequence (Fig. 7B). To further elucidate the molecular mechanism of BAF60c function, we searched for potential association partners of BAF60c. In a yeast two-hybrid screen of a human heart cDNA library, using BAF60c as the bait, we identified few potential interacting factors (BAF155, FEZ1, MYOCD). BAF155 is a component of the BAF complex, which indicates a direct interaction between these two BAF

Table 3. Electrocardiogram analysis of Baf60cMyh6KO mice

<table>
<thead>
<tr>
<th>N=6</th>
<th>Baf60c+/+</th>
<th>Baf60c−/−, Myh6::Cre</th>
<th>Baf60c−/−</th>
<th>Baf60c−/−, Myh6::Cre</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart rate</td>
<td>558±17</td>
<td>560±14</td>
<td>521±11</td>
<td>453±18*</td>
<td>P=0.036</td>
</tr>
<tr>
<td>P duration (ms)</td>
<td>10.2±1.0</td>
<td>7.7±0.7</td>
<td>9.8±0.6</td>
<td>9.2±0.7</td>
<td>NS</td>
</tr>
<tr>
<td>PR duration (ms)</td>
<td>36.5±0.8</td>
<td>36.5±0.5</td>
<td>34.7±0.4</td>
<td>30.6±0.5*</td>
<td>P=0.019</td>
</tr>
<tr>
<td>QRS duration (ms)</td>
<td>7.6±0.2</td>
<td>8.0±0.0</td>
<td>8.2±0.2</td>
<td>11.0±0.5*</td>
<td>P&lt;0.0001</td>
</tr>
<tr>
<td>QT duration (ms)</td>
<td>21.3±0.5</td>
<td>20.0±0.5</td>
<td>20.0±0.6</td>
<td>25.0±0.5*</td>
<td>P=0.031</td>
</tr>
<tr>
<td>QT max duration (ms)</td>
<td>8.7±0.7</td>
<td>8.0±0.0</td>
<td>8.17±0.2</td>
<td>11.0±0.5*</td>
<td>P=0.005</td>
</tr>
<tr>
<td>P height (mV)</td>
<td>0.15±0.02</td>
<td>0.10±0.03</td>
<td>0.13±0.03</td>
<td>0.07±0.01*</td>
<td>P=0.015</td>
</tr>
<tr>
<td>QRS height (mV)</td>
<td>1.1±0.2</td>
<td>1.0±0.2</td>
<td>1.8±0.2</td>
<td>2.2±0.1*</td>
<td>P&lt;0.001</td>
</tr>
</tbody>
</table>

Data are mean±s.e.m. P-value indicates significance assessed by ANOVA. Asterisks indicate significantly different values for individual groups.
complex subunits. Of particular interest among candidate interactors was Myocardin (MYOCD), a transcriptional co-factor of SRF and MEF2c (Creemers et al., 2006; Wang et al., 2001). A GST pull-down assay between GST-fused BAF60c and in vitro synthesized MYOCD confirmed the direct association, and mapped the association domain of MYOCD with BAF60c to amino acids 328–554 (Fig. 7C).

Myl1 is a bona fide direct target of MEF2c/Myocardin (Creemers et al., 2006) and was downregulated in the absence of Baf60c. In an in vitro promoter activation assay, BAF60c could potently enhance the activation of the Myl1 promoter by MYOCD and MEF2c (Fig. 7D). We have not tested a genetic interaction between Baf60c and Myocd. Our data suggest that BAF60c functions as a partner of MYOCD in cardiac development, and that this interaction may be important for the activation of a gene expression program essential for the fundamental functional properties of cardiomyocytes.

DISCUSSION
We showed the requirement for Baf60c in cardiomyocyte function throughout heart development. Loss of Baf60c both prenatally and postnatally resulted in cardiac hypoplasia and defective heart function. Baf60c regulates programs of gene expression that are essential for primary functions of cardiomyocytes, including broad sets of genes essential for sarcomere function and cardiac metabolism.

The Baf60c constitutive knockout phenotype is milder than the mouse shRNA knockdown phenotypes reported earlier (Lickert et al., 2004). The shRNA knockdowns used two independent shRNAs, minimizing the possibility of off-target effects, and the phenotype was rescued by over-expression of BAF60b, indicating significant specificity of the shRNAs. A similar discrepancy exists for Ifitm genes, for which the shRNA phenotype is more severe than that of a genetic deletion (Lange et al., 2008; Tanaka et al., 2005). The possible reasons for the different phenotypes between the shRNA and the genetic null might include effects compounding the loss of Baf60c function, such as overloading of the microRNA processing machinery by overexpressing shRNAs at high levels, other non-specific effects inherent to overexpression of shRNAs in the mouse embryo, or failure to compensate for immediate repression of gene function by RNAi. The genetic deletion here
Fig. 6. See next page for legend.
McArdle, 2007; Nakaseko et al., 2005; Pawar et al., 1999; Tai et al., 2000; Tapscott et al., 1992; You and colleagues, 2008). The changes in cardiac morphology and function observed in Baf60c-deficient embryos indicate that Baf60c is critical for cardiac morphogenesis and function. Therefore, Baf60c is a potential target for therapeutic interventions.

**Materials and Methods**

**ES cell targeting and mouse line establishment**

A Baf60c genomic DNA fragment with loxP sites flanking 1st exon to 4th exon and Frt-Neo-Frt cassettes downstream of 4th exon was constructed using bacterial recombination (Fig. 1A). For gene targeting, 5×10⁶ R1 ES cells were trypsinized and electroporated with 25 µg linearized targeting DNA. The electroporated cells were selected with 160 µg/ml G418 (Gibco #10131) for 7 days. Correctly targeted clones were identified using Southern blots with DNA probes located outside the targeting DNA and labeled with [32P] (Perkin Elmer). The clones were then expanded and used for diploid aggregation. High embryonic stem cell (ESC)-contributed chimera males were bred with ICR and C57/BL6 for germline transmission. Baf60c<sup>los/+</sup> progeny were mated with FLPe-expressing mice [B6.SJ-Tg(ActFLPe)+2930] to remove the Neo cassette between the frt sites and yield Baf60c<sup>los/+</sup> mice. To generate the Baf60c deletion, Baf60c<sup>los/los</sup> mice were mated with pCX-NLS-Cre mice (maintained at the TCP). All animal work was carried out following Canadian Council on Animal Care Guidelines for Use of Animals in Research and Laboratory Animal Care under protocols approved by The Centre for Phenogenomics Animal Care Committee, and in accordance with the guidelines of the University of California, San Francisco (UCSF) Institutional Animal Care and Use Committee (IACUC).

**Mouse and embryo genotyping**

Baf60c<sup>los/+</sup> and Baf60c<sup>los/los</sup> mice were genotyped by PCR using three primers: WT for (5′-GGTCCGAGATTGCCGTA-3′), DEL for (5′-AGCGAGCACCAGCTGTA-3′) and Rev for (5′-CTAGCGAGCTTCGCGATCA-3′). Baf60c deletion band is 250 bp, wild type is 350 bp and Baf60c<sup>los/los</sup> is 470 bp. Postnatal mouse tissues (tail tips or ear notches) and embryo tissues (yolk sac, tails, limb buds) were prepared with the tissue preparation buffer of the Sigma Extract-N-Amp tissue PCR kit (Sigma, XANAT2).

**Histology**

Mouse embryos or tissues were fixed with 4% PFA, dehydrated and embedded with paraffin and sectioned into 4-µm sections then mounted on glass slides. The slides were then stained using standard histology protocols.

**Whole-mount in situ hybridization**

Whole-mount in situ hybridization on mouse embryos at E7.5–10.5 was performed according to standard protocols with the Baf60 in situ hybridization probe (Lickert et al., 2004).

**Optical projection tomography**

Optical projection tomography (OPT) was performed as described (Sharpe et al., 2002) with an OPT system built in-house. E12.5 embryos were harvested, genotyped, fixed with 4% PFA overnight and washed with PBS. The specimens were then embedded in 1% low-melting-point agarose and subsequently cleared using a 1:2 mixture of benzyl alcohol and benzyl butyl phthalate. The cleared embryos were imaged using a Nikon Eclipse 80i microscope equipped with an SLM-AV contribute to the formation of specific subsets of myocardial cells and may play a role in the regulation of specific gene expression programs.
benzoate. The index-matched specimen was suspended from a stepper motor and immersed in a benzyl benzoate bath encompassed in a glass cuvette. Light from a mercury lamp was directed onto the specimen and filter sets were used to create fluorescent images of the specimen. An autofluorescence projection was captured with using a GFP filter set in the illumination and detection light path. Images of the specimen were formed using a Qioptiq Telecentric Zoom 100 microscope equipped with a 0.5× OPTEM objective lens. Projection images were acquired with a Retiga-

![Image of experiments and results](image)

**Fig. 7. Interaction between BAF60c and cardiac transcription factors.** (A) GST-fused Baf60c associates with 35S-labeled, in-vitro synthesized TBX5 and NKX2-5. (B) BAF60c associates with TBX5 through its N-terminal domain. Left: schemes representing serial deletion constructs of BAF60c. Right: mapping the BAF60c-associating domain with GST-fused TBX5. (C) BAF60c associates with full-length Myocardin and serial deletions. Left: schematic representation of Myocardin deletion constructs. The region at amino acids 328–554 of Myocardin was essential for association with Baf60c. (D) BAF60c enhances activation of MYOCD and MEF2c on the My1 luciferase reporter. Data are mean±s.e.m. Asterisks indicate significantly different values (P<0.05 by ANOVA) for individual conditions by t-test.
4000DC CCD camera with pixel size equal to 7.4 μm/pixel. The sample was rotated in finite steps, 0.3°, through a complete revolution totaling 1200 projections. Image reconstruction into a 3D data set was then executed by a modified Feldkamp algorithm in supplied software by SkyScan (NRecon, Bruker, Kontich, Belgium). The resultant OPT images have an isotropic 8.8 micron pixel size.

**RNA-seq**

Mouse embryos from Baf60c<sup>−/−</sup> intercross timed pregnancy at E10.5 and E12.5, or ventricles from Myh6::Cre::Baf60c<sup>−/−</sup> X Baf60c<sup>−/−</sup> intercrosses were harvested. Both males and females were used. Their hearts were individually dissected and snap-frozen with liquid nitrogen. RNA was prepared from each single heart with the PicoPure RNA Isolation kit (Arcturus, Thermo Fisher, Waltham, MA, USA). RNA quantity and quality was analyzed using Agilent RNA 6000 Nano Kit. RNA-seq was performed as described (Christodoulou et al., 2011, 2014). RNA reads were aligned with TopHat/Bowtie (http://ccb.jhu.edu/software/tophat/index.shtml) and Useq (http://useq.sourceforge.net/) was used for the analysis of differential expression. RNAs that showed significant differential expression between wild type and Baf60c<sup>−/−</sup> were selected for analysis, avoiding duplicate and redundant entries.

**Transmission electron microscopy (TEM)**

Mouse E10.5, E12.5 embryonic hearts and 8-week-old adult hearts (males and females) were dissected. For embryonic hearts, the whole heart was used for fixation and section. For adult hearts, pieces of 3–4 mm in size cut from the left ventricle were used as specimens. Pieces of specimen were fixed in a fixative containing 4% formaldehyde and 1% glutaraldehyde in phosphate buffer, pH 7.3, and then post fixed in 1% osmium tetroxide. The specimens were then dehydrated in a graded series of acetone from 50% to 100% and subsequently infiltrated and embedded in Epon-Araldite epoxy resin. The processing steps from post fixation to polymerization of resin blocks were carried out in a microwave oven, Pelco BioWave 34770 (Pelco International, Redding, CA, USA) using similar procedures, with slight modification, as recommended by the manufacturer. Ultrathin sections were cut with a diamond knife on the Reichert Ultracut E (Leica, Vienna, Austria). Sections were stained with uranyl acetate and lead citrate before being examined in the JEM-1011 (JEOL USA, Peabody, MA, USA). Digital electron micrographs were acquired directly with a 1024 X 1024 pixels CCD camera system (AMT, Danvers, MA, USA) attached to the TEM.

**Echocardiography assessment of cardiac functions**

E13.5 embryos were analyzed with a Vevo770 ultrasound machine (VisualSonics, Toronto, Canada). Pregnant Baf60c<sup>−/−</sup> female mice carrying the embryos at the required developmental stages were examined under isoflurane anesthesia. Uteruses were exposed from the incision and scanned with a 30 MHz transducer as described (Lickert et al., 2004). To minimize potential impairment of embryonic physiology, only two or three embryos were scanned for each female, taking about 1 h. The mother’s heart rate was monitored throughout the scanning. For each embryo, the blood flow speed near the mitral and tricuspid valves and aorta was recorded at B-mode. The depth of ventricle walls and ventricle septation was measured at M-mode. After scanning, the embryos were harvested and genotyped. Four to five embryos of each genotype were measured. Adult mice were analyzed using a Vevo2100 ultrasound machine (VisualSonics). The 7–8-week-old animals were anesthetized and scanned with a 30 MHz transducer as described (Zhou et al., 2005). E and A peaks in the left ventricle were measured at B-mode. The chamber dimensions and ventricle wall depths as well as ventricle septation depth were measured at M-mode. For each genotype, five or six mice were measured.

**Electrocardiography**

Mice (male, 8 weeks old) were anesthetized with 1–2% isoflurane, and a lead II ECG was recorded from needle electrodes inserted subcutaneously into the right forelimb and into each hind limb. The signal was recorded for ~1 min. The ECG was recorded with Power Lab/4SP (AD Instruments, Dunedin, New Zealand) and analyzed using the SAECG (signal-averaged electrocardiogram) extension for Chart 4 (v4.2.3 for Macintosh, AD Instruments).

**Immunofluorescence microscopy**

Sarcomeric architecture and organization were assessed in E12.5 and adult hearts via double immunofluorescence staining. Heart tissue was embedded in Tissue-Tek Optimum Cutting Temperature (OCT) compound (Sakura Finetek) and immediately frozen in 2-methylbutane precooled in liquid nitrogen. 5-mm cryosections were mounted on gelatin-coated glass coverslips. Tissue sections were fixed in 4% paraformaldehyde, permeabilized with 0.2% Triton-X 100/PBS and blocked with 2% BSA/1% normal donkey serum/PBS prior to incubation with antibodies. The primary antibodies included: rabbit polyclonal anti-desmin (1:30) (Immunobioscience RP-4023-04), mouse monoclonal anti-sarcomeric α-actinin (1:1000) (Clone EA-53; Sigma A7811), and mouse monoclonal anti-desmoplakin 1/2 (1:1000) (Clone DP-2.15; AbDSerotec 2722-5204) antibodies. The secondary antibodies, obtained from Jackson Immunoresearch Laboratories, included Alexa Fluor 488 goat anti-mouse IgG (1:500), Alexa Fluor 488 goat anti-rabbit IgG (1:500), Texas Red goat anti-mouse IgG (1:500), and Texas Red goat anti-rabbit IgG (1:500). Coverslips were mounted onto slides with Aqua Poly/Mount (Polysciences Inc., Warrington, PA, USA). All sections were analyzed on a DeltaVision RT system with 100× (1.3 NA) objective and a CoolSnap HQ charge-coupled device camera (Photometrics, Tucson, AZ, USA) using softWoRx 3.5.1 software. Images were prepared for presentation using Photoshop CS (Adobe Systems).

**TUNEL analysis**

Cell death on sections was detected using Roche In Situ cell death detection kit Fluorescein (1164795910).

**Yeast two-hybrid assay**

A full-length BAF60c expression construct was used as a bait in a yeast two-hybrid assay conducted by Hybriogenics (www.hybriogenics-services.com/), using a human fetal/adult heart library.

**Luciferase assay**

The Myl11 luciferase construct was as described (Creemers et al., 2006). Combined DNA vectors were transfected into early exponential stage 10T1/2 cells cultured in six-well dishes with Fugene 6 (Roche, 1181443001), following the product manual. After culturing for another 40–48 h, the cells were lysed, and luciferase activity analyzed with Dual-Luciferase Reporter Assay System (Promega E1910). The luciferase activity was normalized with renilla activity. Three biological replicates were prepared for each combination.

**GST-pulldown assay**

<sup>35</sup>S-labeled proteins (TBX5, NKX2-5, RBPjk, NICD, BAF60c serial deletions, Myocardin serial deletions) (Wang et al., 2001) were synthesized with the TnT SP6 coupled reticulocyte lysate system (Promega, L4600) or TnT T7 coupled reticulocyte lysate system (Promega L4610) and labeled with <sup>35</sup>S methionine (Perkin Elmer NEG709A). 5 µl of each synthesized protein was analyzed with SDS-PAGE gel and exposed to X-ray film for evaluation. GST-BAF60c, GST-RBPjk, GST-TBX5 and GST were expressed in E. coli strain BL21 and purified with glutathione Sepharose 4B (GE Healthcare, 17-0756-01). The beads were incubated with <sup>35</sup>S-labeled target proteins overnight at 4°C and washed with PBST for three times. The beads were then boiled in loading buffer. The protein was analyzed with SDS-PAGE gel and autoradiography.

**Statistics**

Data were expressed as mean±s.e.m. Differences among multiple experimental groups were evaluated by ANOVA followed by post hoc Fisher’s LSD test. Pairwise comparisons were evaluated by unpaired two-tailed Student’s t-tests. P<0.05 was considered as significant.

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Competing interests
B.G.B. is a co-founder of Tenaya Therapeutics.

Author contributions

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Data availability
Data are available at GEO (GSE59383) and at https://b2b.bci.utah.edu/gnomex/ (experiment 86R1).

Supplemental data
Supplementary information available online at http://bio.biologists.org/lookup/doi/10.1242/bio.029512.supplemental

References


