Olfactory ensheathing glia are required for embryonic olfactory axon targeting and the migration of gonadotropin-releasing hormone neurons

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Introduction
The anosmia and sterility of Kallmann’s syndrome arise when olfactory axons and gonadotropin-releasing hormone (GnRH) neurons, which are needed for pituitary gonadotropin release, fail to enter the embryonic forebrain (Cadman et al., 2007; Cariboni et al., 2007; Hardelin and Dodé, 2008). GnRH neurons migrate from the embryonic olfactory epithelium along olfactory and/or vomeronasal nerves into the forebrain (Cariboni et al., 2007; Wray, 2010; Wierrman et al., 2011). Recently, spontaneous mutations in the transcription factor gene Sox10 were associated with Kallmann’s syndrome phenotypes: anosmia, hypogonadism and cryptorchidism (Bondurand et al., 2011). Sox10 mutations have been associated with Kallmann’s syndrome phenotypes, but their effect on olfactory system development is unknown. We recently showed that Sox10 is expressed by neural crest-derived olfactory ensheathing cells (OECs). Here, we demonstrate that in homozygous Sox10−/− mouse embryos, OEC differentiation is disrupted; olfactory axons accumulate in the ventromedial olfactory nerve layer and fewer olfactory receptor neurons express the maturation marker OMP (most likely owing to the failure of axonal targeting). Furthermore, GnRH neurons clump together in the periphery and a smaller proportion enters the forebrain. Our data suggest that human Sox10 mutations cause Kallmann’s syndrome by disrupting the differentiation of OECs, which promote embryonic olfactory axon targeting and hence olfactory receptor neuron maturation, and GnRH neuron migration to the forebrain.

Summary
Kallmann’s syndrome is caused by the failure of olfactory axons and gonadotropin-releasing hormone (GnRH) neurons to enter the embryonic forebrain, resulting in anosmia and sterility. Sox10 mutations have been associated with Kallmann’s syndrome phenotypes, but their effect on olfactory system development is unknown. We recently showed that Sox10 is expressed by neural crest-derived olfactory ensheathing cells (OECs). Here, we demonstrate that in homozygous Sox10−/− mouse embryos, OEC differentiation is disrupted; olfactory axons accumulate in the ventromedial olfactory nerve layer and fewer olfactory receptor neurons express the maturation marker OMP (most likely owing to the failure of axonal targeting). Furthermore, GnRH neurons clump together in the periphery and a smaller proportion enters the forebrain. Our data suggest that human Sox10 mutations cause Kallmann’s syndrome by disrupting the differentiation of OECs, which promote embryonic olfactory axon targeting and hence olfactory receptor neuron maturation, and GnRH neuron migration to the forebrain.

Materials and Methods
Embryo collection and sectioning
Sox10−/− mutant mice (Britsch et al., 2001) and wild-type litter-mates of C3HeB/FeJ background were obtained from heterozygous crosses. Embryos were immersion-fixed overnight in 4% paraformaldehyde in phosphate-buffered saline (PBS) at 4°C. Genotypes were determined from tail biopsies as described (Britsch et al., 2001). Embryos were embedded for wax or cryosectioning and sectioned at 5–6 μm (or at 30 μm, for some E16.5 embryos).

Immunohistochemistry
Immunohistochemistry was performed as described (Lassiter et al., 2007). Primary antibodies used were: anti-β galactosidase (chicken, Abcam; 1:1000); anti-BLB (rabbit, Millipore; 1:1000); anti-GnRH-1 (rabbit, Abcam; 1:100); anti-Hu/C/D (mouse IgG2b, Invitrogen; 1:500), anti-laminin (rabbit, Sigma; 1:1000); anti-NPY (rabbit, Abcam, 1:6000), anti-OMP (goat, Wako; 1:500 or 1:1000), anti-p75NTR (rabbit, kind gift of L. Reichardt, University of California at San Francisco, USA; 1:1000), anti-S100 (rabbit, DAKO; 1:50); anti-Sox10 (goat, Santa Cruz Biotechnology; 1:100). Appropriately matched Alexa Fluor 488-, 568- or 594-conjugated secondary antibodies, Alexa Fluor 350-NeutrAvidin and Alexa Fluor 488-streptavidin were obtained from Invitrogen, and biotinylated secondary antibodies from Southern Biotech.

In situ hybridization
Primers against mouse GnRH1 (GenBank accession number NM_008145.2) were designed using Primer3 Input (Rozen and Skalatsky, 2000). Total RNA was extracted

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Key words: Sox10, Olfactory ensheathing glia, GnRH neurons, Kallmann’s syndrome

Research Article

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Fig. 1. See next page for legend.
from the moust and part of the forebrain using Trizol (Invitrogen), and single-strand cDNA generated using Invitrogen’s SuperScript III First-Strand Synthesis System kit. 

Statistical analysis of olfactory receptor neuron maturation and olfactory epithelium thickness

Confocal images covering an optical depth of 15 µm were captured from 30 µm sections through the olfactory mucosa of E16.5 embryos (two wild-type, two Sox10<sup>+/-</sup> and three Sox10<sup>+/+</sup> embryos). Adjacent sections were immunostained for OMP and neuronal βIII tubulin. The region of interest covered a 200 µm length of the nasal septum in the middle portion of the dorsal-ventral span of the olfactory mucosa. Three sections were quantified/embryo for each marker, with each section being 240 µm total rostral-caudal distance; the first section was 300 µm from the most rostral portion of the olfactory bulb. All cells expressing OMP or neuronal βIII tubulin within the imaged regions of interest were counted. For each of the three sections quantified/embryo, the number of OMP-positive and neuronal βIII tubulin-positive cells within the olfactory epithelium on each side of the nasal septum was determined (i.e., 6 measurements/embryo for each marker), and the thickness of the epithelium (from the nasal surface to the basal lamina) measured at three different positions on each side of the septum (i.e., 18 measurements per embryo). The mean/embryo was determined for each measurement, which was converted from pixels to µm and presented as OMP-positive or neuronal βIII tubulin-positive cell/100 µm of olfactory epithelium, or thickness of olfactory epithelium in µm. GraphPad Prism (GraphPad Software, La Jolla, California, USA) was used to perform one-way ANOVA using Tukey’s multiple comparison test (comparing every mean with each other mean) and unpaired 2-tailed t-tests.

Statistical analysis of GnRH neuron distribution

GnRH neurons were counted on 5–6 µm serial sections (10 slides/series: on each slide, each section was collected every 50–60 µm) processed for immunohistochemistry or in situ hybridization to detect GnRH. At least 100 GnRH neurons/embryo were assessed on serial parasagittal sections of E14.5 embryos from three different litters (3 wild-type, 4 heterozygous Sox10<sup>+/+</sup> embryos, 3 homozygous Sox10<sup>+/+/-</sup> embryos) and on serial coronal sections of E16.5 embryos from four different litters (4 wild-type, 4 heterozygous Sox10<sup>+/+/-</sup> embryos, 4 homozygous Sox10<sup>+/+/-</sup> embryos). Differences between the means for groups of the same stage (wild-type versus heterozygous Sox10<sup>+/+/-</sup> embryos, and wild-type versus homozygous Sox10<sup>+/+/-</sup> embryos) were assessed via one-way ANOVA using Dunnett’s multiple comparison test, performed using GraphPad Prism (GraphPad Software, La Jolla, California, USA).

Results and Discussion

Sox10 expression in the developing olfactory system is restricted to OECs (and, at later stages, Bowman’s gland/duct cells)

We aimed to understand how Kallmann’s syndrome phenotypes could result from Sox10 mutations (Bondurand et al., 2007; Barnett et al., 2009). We used in situ hybridization (ISH) and immunostaining to examine Sox10 expression during mouse olfactory system development from E10.5 to neonatal stages (Fig. 1A–O<sup>1</sup>). Our results confirm and extend previous reports (Barraud et al., 2010; Forni et al., 2011) showing that Sox10 expression is restricted to OECs (which are found along the entire length of the olfactory nerve throughout its development), apart from Bowman’s gland/duct cells in the olfactory epithelium at later stages (as we previously described for avian embryos; Barraud et al., 2010). Sox10 expression was not seen (by either ISH or immunostaining) in neurons in the olfactory epithelium at any stage examined (Fig. 1A–O<sup>1</sup>). Likewise, Sox10 expression was not seen in neurons in the vomeronasal organ epithelium (e.g. Fig. 1C–D<sup>1</sup>G–J<sup>1</sup>), or in the neurons (which include GnRH neurons) migrating along olfactory and/or vomeronasal nerves (e.g. Fig. 1C–J<sup>1</sup>). At E16.5, non-neuronal Sox10-positive cells were clearly visible within the olfactory epithelium (Fig. 1M–M<sup>1</sup>). From E17.5 until at least neonatal stages, these were found in large clusters protruding into the mesenchyme (Fig. 1N–O<sup>1</sup>), and as strands projecting across the width of the epithelium (Fig. 1O<sup>1</sup>O<sup>1</sup>). As we previously reported for avian embryos [figure S8 in Barraud et al. (Barraud et al., 2010)], these non-neuronal Sox10-positive cells in the olfactory epithelium can be identified as developing Bowman’s gland/duct cells, which start to protrude from the mouse olfactory epithelium at E17.5 (Cuschieri and Bannister, 1975). Overall, therefore, while GnRH neurons are migrating (Cariboni et al., 2007) and olfactory axons reach the olfactory bulb, Sox10 expression is restricted to OECs during mouse olfactory system development.

OEC differentiation is disrupted after Sox10 deletion

We examined OEC and olfactory system development in mouse embryos in which the Sox10 open reading frame was replaced by lacZ (Britsch et al., 2001). We could identify surviving Sox10-mutant cells by immunostaining for β-galactosidase (β-gal). As reported for other peripheral nerves (Britsch et al., 2001; Paratore et al., 2002), neural crest-derived cells colonized the developing olfactory nerve even after Sox10 deletion, since at E10.5–E12.5, at least some β-gal-expressing cells were present in the olfactory nerve in homozygous Sox10<sup>+/</sup>lacZ embryos (n = 3; Fig. 2A–B<sup>1</sup>). By E16.5, β-gal-expressing cells in heterozygous Sox10<sup>+/</sup>lacZ/+ embryos were distributed along the olfactory nerve from the lamina propria to the ONL (n = 3; Fig. 2C–C<sup>1</sup>). In contrast, β-gal-expressing cells were missing from the lamina propria at E16.5 after Sox10 deletion (n = 3; Fig. 2D<sup>1</sup>, though present on the proximal part of the olfactory nerve, near its entry-point into the olfactory bulb, and within the ONL (Fig. 2D<sup>3</sup>). Similarly, immunoreactivity for S100 (Fig. 2E–F<sup>2</sup>) and p75<sup>TR</sup> (Fig. 2G–H<sup>2</sup>) was absent from the lamina propria after Sox10 deletion (S100, Fig. 2F,F<sup>1</sup>; p75<sup>TR</sup>, Fig. 2G,G<sup>1</sup>; 2G<sup>1</sup>, though present in the

Fig. 1. During mouse olfactory system development, Sox10 expression is restricted to OECs (and, at late stages, to Bowman’s gland/duct cells).

(A–B<sup>1</sup>) At E10.5, in situ hybridization (ISH) on parasagittal sections followed by immunostaining for neuronal βIII tubulin (βIII tub) reveals Sox10 expression in non-neuronal cells associated with the “migratory mass” of olfactory axons and migrating neurons emerging from the olfactory epithelium. Sox10 is not expressed by neurons in either the olfactory epithelium or the migratory mass (examples of each are indicated, respectively, by arrowheads and arrows). (C–F) At E11.5, when the developing vomeronasal organ evacuates from the ventromedial olfactory epithelium (Cuschieri and Bannister, 1975), ISH shows (C–E<sup>1</sup>) Sox10 expression in non-neuronal cells (OECs; arrowheads in E<sup>1</sup>,E<sup>1</sup>) associated with olfactory and vomeronasal (adjacent to the vomeronasal organ) nerves, but no Sox10 expression in the neurons within either the olfactory or vomeronasal epithelia; and (F) GnRH<sup>+</sup>-positive cells within the vomeronasal organ epithelium (arrow) and in the region of the vomeronasal nerve (arrowhead; compare with position of neurons in D<sup>1</sup>–E<sup>1</sup>,G–J<sup>1</sup>). At E12.5, ISH on coronal sections followed by immunostaining for βIII tub shows Sox10 expression in non-neuronal cells associated with the olfactory and vomeronasal nerves, but no background expression within either the olfactory or vomeronasal epithelia. (K–L<sup>1</sup>) At E14.5, immunostaining on coronal sections reveals Sox10-positive, p75<sup>TR</sup>-positive OECs surrounding olfactory nerve fascicles in the lamina propria, but no Sox10 expression in neurons in the olfactory epithelium. (M–N<sup>2</sup>) At E16.5 (M–M<sup>2</sup>) and E17.5 (N–N<sup>2</sup>), ISH for Sox10 on coronal sections followed by immunostaining for βIII tub shows non-neuronal Sox10-positive cells in the olfactory epithelium (arrowheads): these are developing Bowman’s gland/duct cells, which begin to protrude from the basal epithelium from E17.5 (Cuschieri and Bannister, 1975). (O<sup>1</sup>O<sup>1</sup>) In neonates, immunostaining for Sox10 and βIII tub on coronal sections shows that Bowman’s gland/duct cells (arrowheads) maintain Sox10 expression after birth. Abbreviations: fb, forebrain; lp, lamina propria; βIII tub, neuronal βIII tubulin; neo, neonatal; ob, olfactory bulb; oe, olfactory epithelium; on, olfactory nerve; vno, vomeronasal organ. Scale bars: 100 µm (C,D,F,G,K); 50 µm (A,H,J,L,M,N,O); 20 µm (R,L); 10 µm (E).
Fig. 2. See next page for legend.
proximal olfactory nerve and ONL (S100, Fig. 2F,F2; p75NTR, Fig. 2G,G2). However, p75NTR is also expressed by undifferentiated neural crest cells in rodents (Stempke and Anderson, 1992; Rao and Anderson, 1997), and expression of the early glial differentiation marker brain lipid binding protein (BLBP; Fig. 3A–D) (Murdoch and Roskams, 2007), which was also absent from the lamina propria after Sox10 deletion (Fig. 3B,B1), was significantly weaker in the proximal olfactory nerve and ONL (Fig. 3D). This suggests that a glial specification/differentiation defect affects most neural crest cells that colonize the olfactory nerve. Furthermore, we were unable to detect immunoreactivity for the inner ONL-specific OEC marker neuropeptide tyrosine (NPY; Fig. 3E–F2) (Ubink et al., 1994; Ubink and Hökfelt, 2000; Au et al., 2002). Together, these data suggest that in the absence of Sox10, neural crest cells colonize the developing olfactory nerve but normal OEC differentiation fails.

**Sox10 deletion disrupts olfactory axon targeting and olfactory receptor neuron maturation**

The absence of lamina propria OECs at E16.5 in homozygous Sox10lacZ/lacZ embryos was associated with defasciculation of olfactory axon bundles and inappropriate migration of axons within the lamina propria (Fig. 4A–B1). We also noticed an apparent reduction in the number of olfactory receptor neurons (ORNs) expressing the maturation marker olfactory marker protein (OMP) (compare Fig. 4A1,B1; Fig. 4C–E). To investigate this further, we calculated the mean/embryo (± standard error of the mean, s.e.m.) of OMP-positive cells and neuronal βIII tubulin-positive neurons/100 μm of olfactory epithelium, and the thickness of the olfactory epithelium. One-way analysis of variance (ANOVA) using Tukey’s multiple comparison test showed no significant difference for any measurement between wild-type (n = 2) and heterozygous Sox10lacZ/+ embryos (n = 2), so we combined wild-type and heterozygote data (n = 4) for comparison with homozygotes (n = 3). We confirmed that the mean/embryo (± s.e.m.) of OMP-positive cells/100 μm of epithelium (Fig. 4F) was significantly lower for homozygous Sox10lacZ/lacZ embryos (5.65 ± 0.17; n = 3) than for wild-type/heterozygote embryos (10.86 ± 0.69; n = 4) (unpaired 2-tailed t-test: P = 0.0014; t = 6.370; 5 degrees of freedom). In contrast, Sox10 deletion did not affect the mean overall number of neurons/100 μm of epithelium (Fig. 4G: wild-type/heterozygotes: 48.13 ± 1.94; n = 4; homozygotes: 48.71 ± 2.82, n = 3), or the mean thickness of the olfactory epithelium (Fig. 4H: wild-type/heterozygotes: 73.87 ± 1.91 μm; n = 4; homozygotes: 74.60 ± 2.13 μm; n = 3), suggesting that Sox10 deletion specifically affects ORN maturation.

![Fig. 2. After Sox10 deletion, neural crest cells colonize the olfactory nerve but are missing from the lamina propria by E16.5. (A–B1)](image)

In homozygous Sox10lacZ/lacZ embryos at E11.5 (A–A1) and E12.5 (B–B1), immunostaining on parasagittal sections reveals β-gal-positive cells associated with the olfactory nerve (arrows, A2,B2). β-gal expression in mesenchymal cells beneath the olfactory epithelium (B1) may reflect protein perdurance in neural crest-derived cells. (C–D1) At E16.5, immunostaining on coronal sections shows β-gal-positive cells (arrows) throughout the olfactory nerve from the lamina propria to the ONL in heterozygous Sox10lacZ/+ embryos (C–C1), but absent from the lamina propria after Sox10 deletion (D–D1). Arrowheads in C1,D1 indicate β-gal-positive prospective Bowman’s gland duct cells in the epithelium, which express Sox10 (Fig. 1N–O1). Fainter β-gal immunoreactivity in the cribriform plate and nasal septum cartilage (C1,D1,D2) may be specific: weak β-gal staining was previously reported in limb cartilage condensations in Sox10lacZ embryos (Britsch et al., 2001). (E–F2) At E16.5, immunostaining on coronal sections shows p75NTR expression (arrows) throughout the olfactory nerve from the lamina propria to the ONL in wild-type embryos (E–F2), but absent from the lamina propria after Sox10 deletion (F–F2). (G–H1) At E16.5, immunostaining on coronal sections shows λ-gal-positive cells (arrows) throughout the olfactory nerve from the lamina propria to the ONL in wild-type embryos (G–H1), but absent from the lamina propria after Sox10 deletion (H–H1). Abbreviations: β-gal, β-galactosidase; cp, cribriform plate; fb, forebrain; lp, lamina propria; n33thb, neuronal βIII tubulin; ns, nasal septum; ob, olfactory bulb; oe, olfactory epithelium; on, olfactory nerve; onl, olfactory nerve layer; vn, vomeronasal nerve; vno, vomeronasal organ. Scale bars: 100 μm (A,B,C,D,E,F,G,H); 50 μm (A1,B1,C1,D1,D2,E1,F1,F2,G1,G2,H1,H2); 25 μm (A2,A3,B2,B3).
Fig. 3. After Sox10 deletion, OEC differentiation is defective. All images show E16.5 coronal sections. (A–B1) Expression of the early glial differentiation marker BLBP is seen in OECs ensheathing olfactory nerve fascicles (labelled by immunostaining for olfactory marker protein) in the wild-type lamina propria (A,A1), but not after Sox10 deletion (B,B1). (C,D) BLBP expression in the ONL is much stronger in wild-type (C) than homozygous Sox10lacZ/lacZ embryos (D). (E–F2) At E16.5, NPY expression is seen in the inner ONL of wild-type embryos (arrows, E1,E2) but undetectable after Sox10 deletion (F1,F2). Abbreviations: epl, external plexiform layer; lp, lamina propria; ob, olfactory bulb; OMP, olfactory marker protein; on, olfactory nerve; onl, olfactory nerve layer; onl-i, inner olfactory nerve layer; onl-o, outer olfactory nerve layer. Scale bars: 100 μm (E,E1,F,F1), 50 μm (C,D), 25 μm (E1 inset), 10 μm (A,B).
Fig. 4. Sox10 deletion disrupts olfactory axon targeting and ORN maturation. All images show E16.5 coronal sections. (A–E) Olfactory mucosa sections immunostained for the maturation marker OMP plus (A–B1) NCAM or (C–E) neuronal βIII tubulin. Relative to wild-type (A,A1,C) or heterozygote embryos (D), homozygous Sox10lacZ/lacZ embryos (B,B1,E) displayed defasciculated olfactory nerve bundles and inappropriately migrating axons within the lamina propria (asterisk in B,B1), and fewer OMP-positive neurons (compare A1,B1; C–E). (F–H) Bar charts showing the mean/embryo + s.e.m. for wild-type/heterozygote embryos (2/genotype) versus homozygous Sox10lacZ/lacZ embryos, of: (F) OMP-positive cells/100 μm of olfactory epithelium (**P=0.0014; 2-tailed unpaired t-test); (G) neuronal βIII tubulin-positive cells/100 μm of epithelium; (H) olfactory epithelial thickness. Sox10 deletion only affects the number of mature (OMP-positive) ORNs. (I,J) Olfactory bulb sections immunostained for the axonal marker NCAM. Relative to wild-type embryos (I), the ONL in homozygous Sox10lacZ/lacZ embryos (J) is much thinner in dorsal and lateral regions of the bulb while the ventromedial ONL is much thicker. (K–L1) NCAM immunostaining showing clearly separated uniform bilateral ONLs in a wild-type embryo (K) versus a merged, ventromedial ONL (asterisk) in a homozygous Sox10lacZ/lacZ embryo (L,L1). Arrowheads in L,L1 highlight axonal whorls/balls. Abbreviations as in Fig. 2. Scale bars: 200 μm (K,L), 100 μm (I,J,L1), 50 μm (A,B,C).
Fig. 5. A significantly smaller proportion of GnRH neurons enters the forebrain after Sox10 deletion. (A) At E12.5, immunostaining on coronal sections for neuronal βIII tubulin and the neuronal RNA-binding protein HuC/D revealed unusually large aggregates of multiple neuronal cell bodies on the vomeronasal nerve in both heterozygous Sox10lacZ/− and homozygous Sox10lacZ/lacZ embryos. (B) Schematic representation of the distribution of GnRH neurons (black spots) at E14.5 on parasagittal sections of 3 wild-type, 3 heterozygous Sox10lacZ/+ and 3 homozygous Sox10lacZ/lacZ embryos. (C) Schematic representation of the distribution of GnRH neurons (black spots) at E16.5 on coronal sections at 3 different rostrocaudal levels of 4 wild-type, 4 heterozygous Sox10lacZ/− and 4 homozygous Sox10lacZ/lacZ embryos. (D) Bar charts showing the mean percentage/embryo ± s.e.m. of GnRH-positive cells that had entered the brain for wild-type, heterozygous Sox10lacZ/− and homozygous Sox10lacZ/lacZ embryos (3 embryos/genotype at E14.5; 4 embryos/genotype at E16.5; ≥100 GnRH neurons counted/embryo). At E14.5, the mean percentage/embryo of GnRH neurons in the brain was significantly lower than wild-type for both heterozygous Sox10lacZ/− and homozygous Sox10lacZ/lacZ embryos (**P<0.01; one-way ANOVA using Dunnett’s multiple comparison test). By E16.5, the mean percentage/embryo of GnRH neurons in the brain was no longer significantly different from wild-type for heterozygous Sox10lacZ/− embryos, but was four-fold lower than wild-type for homozygous Sox10lacZ/lacZ embryos (**P<0.005; one-way ANOVA using Dunnett’s multiple comparison test). (E–F’) Examples of GnRH neurons identified after immunostaining E16.5 coronal sections of wild-type hypothalamus (E,E’) and homozygous Sox10lacZ/lacZ olfactory mucosa (F,F’). Abbreviations as in Fig. 2. Scale bars: 200 μm (E), 100 μm (A, main panels; E’,F), 25 μm (A, insets; F’), 10 μm (E’, insets).
multiple comparison test showed that at E14.5, the mean percentage/embryo of GnRH neurons that had entered the brain was significantly lower (P<0.01) in both heterozygous Sox1{lox/lox}mutants (13.7±0.9%; n=3) and homozygous Sox1{lox/lox}mutants (13.3±4.8%; n=3) than in wild-type embryos (37.7±1.2%; n=3) (Fig. 5D). By E16.5, there was no longer any significant difference between heterozygote (38.8±6.9%; n=4) and wild-type embryos (51.9±4.7%; n=4), but four-fold fewer GnRH neurons were present in the brain in homozygous Sox1{lox/lox}mutants (11.9±3.9%; n=4) than in wild-type embryos (P<0.005; also see Fig. 5D). Examples of GnRH neurons in the hypothalamus of wild-type embryos and the olfactory mucosa of homozygous Sox1{lox/lox}mutants are shown in Fig. 5E-1F. These data suggest that GnRH neuron migration to the forebrain is delayed when one copy of Sox10 is missing, and stalled after Sox10 deletion. A recent report describing a close association between migrating GnRH neurons and OECs (Geller et al., 2013) is consistent with the important role for OECs in GnRH neuron migration that we have described here.

We conclude that human Sox10 mutations cause Kallmann’s syndrome phenotypes (Bondurand et al., 2007; Barnett et al., 2009) by disrupting the differentiation of OECs, which, as shown here, promote olfactory axon targeting, ORN maturation (most likely because of their importance for olfactory axon targeting) and GnRH neuron migration. A neural crest defect in Kallmann’s syndrome is supported by its inclusion within CHARGE syndrome (Pinto et al., 2005), an autosomal dominant disorder caused by deletions at the CHD7 gene locus cause Waardenburg syndrome types 2 and 4. Am. J. Hum. Genet. 81, 1169-1185.

Note added in proof

While our manuscript was in revision, another study was published showing that loss-of-function mutations in SOX10 cause Kallmann’s syndrome with deafness and describing the role for OECs in GnRH neuron migration that we have described here.

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Author Contributions

P.B. and C.V.H.B. conceived the project, designed the experiments and wrote the manuscript. P.B. and J.A.S. performed the experiments and discussed and interpreted the data with C.V.H.B. C.C.S. and M.W. provided embryos, reagents and expertise.

Competing Interests

The authors have no competing interests to declare.

References


