A proteomic study of mitotic phase-specific interactors of EB1 reveals a role for SXIP-mediated protein interactions in anaphase onset

Naoka Tamura1, Judith E. Simon1,2, Arnab Nayak1,3, Rajesh Shenoy1, Noriko Hiroi4, Viviane Boilot1, Akira Funahashi4 and Viji M. Draviam1,*

ABSTRACT

Microtubules execute diverse mitotic events that are spatially and temporally separated; the underlying regulation is poorly understood. By combining drug treatments, large-scale immunoprecipitation and mass spectrometry, we report the first comprehensive map of mitotic phase-specific protein interactions of the microtubule-end binding protein, EB1. EB1 interacts with some, but not all, of its partners throughout mitosis. We show that the interaction of EB1 with Astrin-SKAP complex, a key regulator of chromosome segregation, is enhanced during prometaphase, compared to anaphase. We find that EB1 and EB3, another EB family member, can interact directly with SKAP, in an SXIP-motif dependent manner. Using an SXIP defective mutant that cannot interact with EB, we uncover two distinct pools of SKAP at spindle microtubules and kinetochores. We demonstrate the importance of SKAP’s SXIP-motif in controlling microtubule growth rates and anaphase onset, without grossly disrupting spindle function. Thus, we provide the first comprehensive map of temporal changes in EB1 interactors during mitosis and highlight the importance of EB protein interactions in ensuring normal mitosis.

KEY WORDS: Cell cortex, Kinetochore, Microtubule, Mitosis, Plus-end

INTRODUCTION

During mitosis, microtubules play a crucial role in multiple concurrent events: in early mitosis, microtubules assemble a bipolar spindle, capture chromosomes at specialized sites called kinetochores, power chromosome movements and rotate the bulky spindle apparatus towards a predetermined axis. Following the completion of chromosome congression in metaphase, cells initiate anaphase – a phase when microtubules pull and separate chromatids apart, establish the plane for cleavage furrow formation and facilitate anaphase cell elongation. How mitotic microtubules are controlled to coordinate such diverse tasks, in a spatially and temporally defined manner, is a fascinating and poorly understood biological problem.

The microtubule-end binding protein EB1 forms a comet-like structure specifically at the plus-ends of growing microtubules (Mimori-Kiyosue et al., 2000) and a crescent-like structure at the kinetochore following microtubule attachment (Tirnauer et al., 2002). Although members of the EB family, EB1 and EB3, act redundantly to regulate microtubule growth in interphase cells (Komarova et al., 2009; Komarova et al., 2005), they control diverse spatially and temporally separated mitotic events, including kinetochore alignment (Draviam et al., 2006; Green et al., 2005), spindle orientation (Brüning-Richardson et al., 2011; Draviam et al., 2006; Ferreira et al., 2013; Green et al., 2005; Toyoshima and Nishida, 2007) and post-cytokinetic cell spreading (Ferreira et al., 2013). However, the underlying molecular mechanisms are not understood.

Several EB1 or EB3 interacting proteins have been identified using large-scale immunoprecipitations from asynchronous cell populations (Berrueta et al., 1999; Geraldo et al., 2008; Gu et al., 2006; Meireles et al., 2011; Rogers et al., 2004; Schroder et al., 2011). In interphase, EB1 interacts with several partners to modulate interphase microtubule plus-end function (reviewed in Akhmanova and Steinmetz, 2008; Jiang et al., 2012). A clear molecular understanding of EB1’s mitotic function requires a comprehensive list of EB-interactors from temporally separated distinct phases of mitosis and this is currently lacking.

EB proteins interact with several partner proteins bearing either a CAP-Gly rich domain or an S/T-X-I/L-P motif (reviewed as ‘SXIP-motif’) (reviewed in Akhmanova and Steinmetz, 2010; Tamura and Draviam, 2012). While residues adjacent to the SXIP motif could render further specificity for EB1 protein interactions (Buey et al., 2011; Honnappa et al., 2009; Jiang et al., 2012), phosphorylation around EB1 binding motifs is reported to be a crucial determinant of EB1 protein interactions during both interphase and mitosis (Honnappa et al., 2005; Smyth et al., 2012; Kumar et al., 2012; Kumar et al., 2009; van der Vaart et al., 2011; Watanabe et al., 2009; Wittmann and Waterman-Storer, 2005; Zimniak et al., 2009). However, it is not known if disruption of EB protein interaction would modulate microtubule dynamics during mitosis as it does during interphase (reviewed in Tamura and Draviam, 2012).

To determine how microtubule plus-ends execute distinct mitosis phase-specific events, we searched for EB1 interactors from two distinct phases of mitosis using large-scale immunoprecipitation and mass spectrometry. Our proteome-wide effort revealed the spindle and kinetochore associated protein, SKAP, as a mitotic phase dependent interactor of EB1.

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We show that SKAP’s SXIP-motif is essential for interacting with both EB1 and EB3. Using an SXIP defective mutant, we show that the SXIP-motif is important for proper mitotic microtubule growth rates and SKAP overexpression induced delay in anaphase onset. Our findings show that an excess of SKAP-EB interaction can result in an anaphase onset delay, without grossly affecting other microtubule-mediated functions such as bipolar assembly or chromosome congression. We present a model wherein finely regulated interaction of microtubule plus-end complexes is a key rate-limiting factor for determining the onset of anaphase.

RESULTS
Mitotic phase determined interactions of EB1
To identify microtubule plus-end bound complexes from distinct phases of mitosis, we performed large-scale immunoprecipitation of Flag-tagged EB1 (Flag-EB1) from prometaphase and anaphase cell lysates. Flag-tagged Nuf2 (Flag-Nuf2) was used as a bait control because the human Ndc80-Nuf2 complex is a core-kinetochore protein which was shown using Electron Microscopy to interact with microtubule walls (Cheeseman et al., 2006) and to associate with disassembling microtubule-ends (Umbreit et al., 2012), as opposed to EB1 that binds selectively to growing microtubule-ends (Komarova et al., 2009; Mimori-Kiyosue et al., 2000).

For large-scale enrichment of mitotic cells, we treated cells with DMA, an Eg5 inhibitor that induces monopolar spindles and arrests cells in prometaphase, and then performed mitotic shakeoff for isolating rounded-up prometaphase cells; for anaphase cells, we washed the rounded-up cells to remove the inhibitor and synchronously released the cells into anaphase (Fig. 1A). DMA treatment of UTA6-Flag-EB1 cells allowed the enrichment of mitotic cells to approximately 50% of the total cell population (data not shown). As expected from our previous studies of monopolar to bipolar spindle conversion (Shrestha et al., 2014), prometaphase UTA6 cells were predominantly in anaphase following a 45 min release from DMA treatment, as confirmed using microscopy of UTA6 cultures (supplementary material Fig. S1A). Thus, the DMA treatment and mitotic shakeoff protocol allowed us to obtain high-quality lysates of cells from two different mitotic phases.

To exclude immunoprecipitation artefacts, we performed five quality control steps: First, we ensured that Flag-EB1 was expressed at levels comparable to endogenous EB1 by modulating Tetracycline-release induced protein expression conditions (Fig. 1B). Second, we confirmed that the localization of Flag-EB1 was restricted to the plus-ends of microtubules (Fig. 1C,D). Third, we used Flag peptides to specifically elute Flag-EB1 and associated complexes. Fourth, to exclude contaminants, we compared mass spectrometry data of immunoprecipitates from UTA6-Flag-EB1 and UTA6-Flag-Nuf2 cells and excluded all common interactors. Although we may lose their common interactors at the kinetochore, we were not concerned because first, Gene Ontology (GO) analysis of the excluded common interactors did not show any enrichment for microtubule plus-end complexes and second, we did not find EB proteins in any of the Nuf2 immunoprecipitates (data not shown). Finally, to ensure that the protein candidates identified are true hits in the proteomic database, we only considered proteins that were identified using at least two distinct peptides. We first confirmed that these five stringent steps allowed us to immunoprecipitate established EB1 interactors, such as p150 (DCTN1), specifically from UTA6-Flag-EB1 but not UTA6-Flag-Nuf2 cell line (Fig. 1E).

For building the EB1-interacting proteome from distinct mitotic phases, we extracted from our mass spectrometry data only the specific interactors of EB1 that were reproducibly found in at least 2 repeats of Flag-EB1 immunoprecipitations, and never in any of the repeats of Flag-Nuf2 immunoprecipitations (Tables 1, 2). To exclude artefacts, we considered only those proteins where at least two distinct peptides could be recovered. Some interactors of EB1 are common to both prometaphase and anaphase; however, many others are specific to either prometaphase or anaphase (Fig. 1F; supplementary material Fig. 1B). We then compared our list of mitotic phase-specific flag-EB1 interactors against previously obtained list of GST-EB1 interactors from asynchronous human cell cultures that should include a small proportion of mitotic cells (Jiang et al., 2012). This comparison study showed that while nearly half of our hits could be observed in asynchronous conditions as well, at least 40% of the hits could be visualized only in conditions that enrich for mitotic cells (Tables 1, 2).

A strong evidence of success in our proteome-wide search for mitosis phase-specific plus-end complexes, we found the plus-end tracking kinesin, Kif18b (KIF18B) from prometaphase, but not anaphase, extracts of UTA6-Flag-EB1 cells (Fig. 1F; supplementary material Fig. 1B). This is consistent with previous studies showing the plus-end localization of Kif18B in early mitosis but not anaphase (Lee et al., 2010; Stout et al., 2011). Thus, our data presents the first comprehensive map of mitotic phase-specific interactors of EB1.

Astrin-SKAP complex is a mitotic phase-specific interactor of EB1
Among the protein complexes that were immunoprecipitated with EB1, the Astrin (SPAG5)-SKAP complex, a known regulator of spindle and kinetochore function (Dunsch et al., 2011; Gruber et al., 2002; Schmidt et al., 2010) and a marker of kinetochores bearing mature attachments to microtubule-ends (Shrestha and Draviam, 2013), was reproducibly observed in Flag-EB1 immunoprecipitates from prometaphase cells but not anaphase cells (Fig. 1F; supplementary material Fig. 1B).

Interaction between EB1 and SKAP is known (Wang et al., 2012), but whether the interaction is subjected to mitotic phase dependent regulation was not known. Therefore, to confirm our findings from the proteome-wide study, we investigated if EB1 interacts with SKAP in a mitotic phase-specific manner using quantitative fluorescent immunoblotting. This allowed us to quantify and compare the amount of SKAP across three independent repeats of Flag-EB1 immunoprecipitations from prometaphase and anaphase cells (Fig. 2A,B). Anti-Flag immunoprecipitations from UTA6 Flag-EB1 cell lysates reproducibly showed that the interaction of EB1 with SKAP was on average four-fold higher in prometaphase lysates with high Cyclin-B levels, compared to anaphase lysates with low Cyclin-B levels (Fig. 2B; supplementary material Fig. S1C). Because Cyclin-B starts degrading at the end of metaphase and continues into anaphase but is not degraded in prometaphase (Clute and Pines, 1999), the difference in Cyclin-B levels further confirms the successful separation of the two mitotic phases. These data reveal the Astrin-SKAP complex as a mitotic phase determined interactor of EB1.

Both EB1 and EB3 interact directly with SKAP of the Astrin-SKAP complex
EB1 and EB3 have a highly conserved EB homology domain (reviewed in Tamura and Draviam, 2012). However, during
mitosis, the two proteins play non-redundant roles. EB1 is required for spindle positioning in metaphase and chromosome segregation in anaphase (Draviam et al., 2006). In contrast, EB3, but not EB1, is required for spindle positioning in anaphase (Ferreira et al., 2013). It is not known if both EB1 and EB3 are capable of interacting with SKAP. Although many interphase partners of EB1 can redundantly bind to EB3 as well (Bu and Su, 2003; Komarova et al., 2005; Mimori-Kiyosue et al., 2005; van der Vaart et al., 2011), it is not known if mitotic interactors have similar redundancy, and this is important to study because EB1 and EB3 regulate non-redundant mitotic functions (Ferreira et al., 2013).

To test if both EB1 and EB3 are capable of interacting with SKAP, we used yeast two-hybrid (Y2H) assays (Fig. 2C,D). For control studies, we mutated LP into NN in the S/T-X-I/L-P motif since similar mutations have been shown to abrogate the interaction between EB and its partners (Honnappa et al., 2009; Jiang et al., 2012). Y2H studies showed that EB1 interacts with SKAP in mitosis, the two proteins play non-redundant roles. EB1 is required for spindle positioning in metaphase and chromosome segregation in anaphase (Draviam et al., 2006). In contrast, EB3, but not EB1, is required for spindle positioning in anaphase (Ferreira et al., 2013). It is not known if both EB1 and EB3 are capable of interacting with SKAP. Although many interphase partners of EB1 can redundantly bind to EB3 as well (Bu and Su, 2003; Komarova et al., 2005; Mimori-Kiyosue et al., 2005; van der Vaart et al., 2011), it is not known if mitotic interactors have similar redundancy, and this is important to study because EB1 and EB3 regulate non-redundant mitotic functions (Ferreira et al., 2013).

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Table 1. Summary of mass spectrometry based identification of EB1 interactors in DMA-released mitotic cell extracts

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<th>Protein ID</th>
<th>Protein description</th>
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SKAP Wild-Type (WT) but not the SKAP(NN) mutant with a defective SXIP-motif (Fig. 2C), consistent with previous report (Wang et al., 2012), confirming the role of the SXIP-motif in mediating SKAP-EB1 interaction. In addition, we found that similar to EB1, EB3 can also interact directly with SKAP and this interaction is also dependent on SKAP’s SXIP-motif (Fig. 2D). We conclude that both EB1 and EB3 are capable of interacting with SKAP, in an SXIP-motif dependent manner. Thus, SKAP joins a family of proteins that are capable of interacting with either EB1 or EB3 (Bu and Su, 2003; Komarova et al., 2005; Mimori-Kiyosue et al., 2005; van der Vaart et al., 2011), unlike others that selectively interact with only one of the EB proteins (Goldspink et al., 2013; Straube and Merdes, 2007).

We next tested if Astrin, the other member of the Astrin-SKAP complex directly interacted with EB1, since Astrin is also immunoprecipitated with Flag-EB1 in a mitosis phase-specific manner (Tables 1, 2). Our Y2H studies showed that Astrin did not interact with EB1 although as expected Astrin interacted with SKAP (supplementary material Fig. S2). This shows that EB1 interacts specifically with SKAP of the Astrin-SKAP complex.

An evolutionarily conserved Aurora B-consensus site exists proximal to the SXIP-motif of SKAP (Fig. 2E). Previous studies have shown that the interaction between EB and its partners can be negatively regulated by phosphorylation close to the SXIP-motif (Buey et al., 2012) and such negative regulation has been reported during mitosis (Kumar et al., 2012; Zimniak et al., 2009). Moreover, Aurora-B is known to negatively regulate SKAP recruitment to kinetochores (Schmidt et al., 2010). Therefore, we investigated if SKAP-EB interaction is controlled by phosphorylation of the Aurora-B consensus site proximal to the SXIP-motif using either non-phosphorylatable (phospho-dead) or phospho-mimetic mutants by mutating RAT108 into RAA or RAE, respectively. Y2H studies showed that phospho-mimetic mutation of SKAP at T108 (RAT to RAE) significantly reduces SKAP interaction with EB1 (Fig. 2C,G) and completely abolishes SKAP interaction with EB3 (Fig. 2D,F). In contrast, the SKAP (RAT to RAA) mutant was able to interact with both EB1 and EB3 (Fig. 2C,D), showing the role of charged residues in modulating EB-SKAP interactions. Thus, in addition to the SXIP-motif, electrostatic interactions surrounding the SXIP-motif are critical for SKAP-EB interaction.

**SXIP-motif is essential for SKAP recruitment to spindle microtubules but not kinetochores**

The functional significance of SKAP-EB interaction in vivo is not known, although in vitro studies show SKAP’s interaction with EB1 to be important for its microtubule plus-end loading (Wang et al., 2012).

To address the role of SKAP-EB interaction in vivo, we disrupted the interaction in cells using the SKAP(NN) mutant. We established HeLa FRT/TO cell lines that conditionally expressed either GFP-tagged SKAP(WT) or GFP-SKAP(NN) in the presence of Tetracycline. Using immunofluorescence, we first analysed SKAP localization at kinetochore, spindle poles and spindle microtubules in cells treated with MG132, a proteasome inhibitor that arrests cells in metaphase (Fig. 3A). Following MG132 treatment, cells expressing GFP-SKAP(WT) or GFP-SKAP(NN) mutant displayed congressed chromosomes and both proteins localized normally to spindle poles and kinetochores (Fig. 3B,C). However, fluorescence intensity of GFP-SKAP(NN) mutant on spindle microtubules was much reduced compared to GFP-SKAP(WT).
### Table 2. Summary of mass spectrometry based identification of EB1 interactors in DMA-arrested released mitotic cell extracts

<table>
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<th>Protein ID</th>
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<tr>
<td>Q9Y448</td>
<td>Putative TRAF4-associated factor 1 (KNSTRN)</td>
<td>2</td>
<td>35,701</td>
<td>95</td>
<td>3</td>
<td>8.5</td>
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<td>4</td>
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<td>Guanine nucleotide-binding protein G(k) subunit alpha (GNAI3)</td>
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<td>41,076</td>
<td>106</td>
<td>4</td>
<td>7.3</td>
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<td>O15020</td>
<td>Spectrin beta chain, brain 2 (SPTBN2)</td>
<td>2</td>
<td>272,526</td>
<td>120</td>
<td>5</td>
<td>1.4</td>
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<tr>
<td>Q92828</td>
<td>Coronin-2A (CORO2A)</td>
<td>2</td>
<td>60,239</td>
<td>67</td>
<td>1</td>
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<td>71</td>
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<td>5.2</td>
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<tr>
<td>P11233</td>
<td>Ras-related protein Ral-A (RALA)</td>
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<td>70</td>
<td>1</td>
<td>6.8</td>
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<tr>
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<td>37</td>
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<td>Q8NF91</td>
<td>Nesprin-1 (SYNE1)</td>
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<td>44</td>
<td>3</td>
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<tr>
<td>P11166</td>
<td>Solute carrier family 2, facilitated glucose transporter member 1 (SLC2A1)</td>
<td>2</td>
<td>54,391</td>
<td>34</td>
<td>1</td>
<td>1.6</td>
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Table lists EB1 interactors reproducibly found at least twice in anti-Flag immunoprecipitations from DMA-arrested extracts of UTA6 Flag-EB1 cell line and never in UTA6 Flag-Nuf2 cell line. SKAP is referred to as KNSTRN. See Table 1 for explanation of column headings.
revealing the role of SXIP-motif in localizing SKAP onto spindle microtubules.

We then measured microtubule growth using EB3 – a marker of growing microtubule-ends (Bu and Su, 2001; Nakagawa et al., 2000) – in live-cells co-expressing EB3-mKate and either GFP-SKAP(WT) or GFP-SKAP(NN) mutant. We assessed the instantaneous growth velocities of EB3-mKate comets automatically using the plus-tip tracker software (Matov et al., 2010). Microtubule growth velocities of EB3 comets in mitotic cells expressing EB3-mKate alone (average peak values from plus-tip tracker data: Control: 15 μm/min; n comets = 17,923) was comparable to previously reported values for EB3 (Sironi et al., 2010). Microtubule growth velocities of EB3 comets in mitotic cells expressing EB3-mKate alone (average peak values from plus-tip tracker data: Control: 15 μm/min; n comets = 17,923) was comparable to previously reported values for EB3 (Sironi et al., 2010).
However, cells over-expressing GFP-SKAP(WT), showed a significant reduction in microtubule growth rates compared to control cells (GFP-SKAP(WT): 12 μm/min; ncomets = 3347). Slightly higher average peak values of microtubule growth rates were observed in GFP-SKAP(NN) expressing cells (GFP-SKAP(NN) cells: 17 μm/min; ncomets = 4582), compared to controls. However, the overall distribution of growth velocities was significantly different between control and GFP-SKAP(WT) expressing cells, but not GFP-SKAP(NN) mutant expressing cells (Fig. 3D). We conclude that the regulation of SKAP-EB interaction is important for maintaining normal microtubule growth velocity during mitosis.

Collectively, these data shed first insight into the existence of two distinct pools of SKAP: a spindle microtubule associated pool that influences microtubule growth in an SXIP-motif dependent manner and a kinetochore bound pool that binds to congressed kinetochores in an SXIP-motif independent manner.

SKAP overexpression delays anaphase onset, in an SXIP-motif dependent manner

We next investigated if SKAP-EB interaction is important for mitotic progression using time-lapse microscopy (Fig. 4A). An interesting difference emerged between cells overexpressing SKAP(WT) or SKAP(NN) mutant. Consistent with a previous report of metaphase arrest in SKAP over-expressing cells (Dunsch et al., 2011), our time-lapse studies showed a clear metaphase arrest and delay in anaphase onset in cells overexpressing GFP-SKAP(WT) (Fig. 4B,C). Strikingly, however, cells overexpressing the GFP-SKAP(NN) mutant did not display any delay in anaphase onset, compared to the parental HeLa FRT/TO cell line (Fig. 4D), showing the anaphase delay to be SXIP-motif dependent. We confirmed that this striking difference in anaphase onset times between cells expressing SKAP(WT) or SKAP(NN) is not due to a difference in the amount of GFP-tagged SKAP(WT) and SKAP(NN) (supplementary material Fig. S3A,B). Because SKAP(WT)
overexpressing but not SKAP(NN) cells display a prominent delay in anaphase onset despite normal chromosome congression and bipolar spindle assembly (Fig. 3B). We conclude that excessive SKAP-EB interaction delays anaphase onset without grossly disrupting other microtubule-mediated mitotic functions.

SKAP overexpression induced delay in anaphase onset could arise from either a biochemical inability in transitioning from metaphase to anaphase because of APC/C inactivation or a physical inability in pulling and segregating sister chromatids apart. To test if the delay in anaphase induced by SKAP overexpression is due to a delay in APC/C activation, we performed time-lapse microscopy of cells expressing either GFP-SKAP(WT) or GFP-SKAP(NN) mutant in the presence of NMS-P715, an inhibitor of MPS1 kinase that is required for kinetochore bound checkpoint signaling and subsequent Mitotic Checkpoint Complex assembly (Tipton et al., 2013; Zich et al., 2012). Around 80% of metaphase cells expressing GFP-SKAP(WT) or GFP-SKAP(NN) mutant were observed to initiate anaphase within 50 min of treating cells with MPS1 inhibitor (Fig. 4E). These data show that SKAP overexpression induced delay in anaphase is dependent on the spindle assembly checkpoint-induced inhibition of the APC/C. Thus, SKAP overexpression delays anaphase by inhibiting APC/C activation, in an SXIP-motif dependent manner.

Although overexpression studies are difficult for functional dissection, we find that the SKAP (SXIP) mutant lacks the SKAP (WT) overexpression induced phenotype. This suggests that there is a fine balance in SXIP mediated interactions and upsetting this balance can delay anaphase onset. This data obtained through SKAP overexpression studies is clinically relevant.
because the protein is overexpressed in breast carcinomas (Wright and Brooks, 2013) and recurrent mutations in SKAP are correlated with aneuploidy in squamous cell carcinoma (Lee et al., 2014).

**Co-depletion of EB1 and EB3 delays the onset of anaphase**

Our data thus far shows that the abrogation of SKAP-EB interaction, in SKAP(NN) expressing cells, fully rescues SKAP overexpression induced delay in anaphase onset. Therefore, we
hypothesized that excessive SKAP-EB interaction might lead to a loss of EB function and thus delay anaphase onset. If our hypothesis is correct, we would expect a similar anaphase delay following the loss of EB1 and EB3 proteins. In support of our hypothesis, a recent study indicated that the average anaphase onset is slightly reduced in cells co-depleted of EB1 and EB3 (Ferreira et al., 2013). However, it is not known whether anaphase onset delay is related to chromosome congression delay in cells depleted of EB1 and EB3 proteins. Therefore, we compared the rates of anaphase onset and completion of chromosome congression in EB1 and EB3 depleted HeLaHi2B-GFP; mCherry-Tubulin cells using time-lapse microscopy (Fig. 5A,B). To ensure that both EB1 and EB3 are fully depleted in our time-lapse microscopy studies, we harvested cell extracts at the end of each time-lapse imaging session and quantified the extent of protein depletion using fluorescent immunoblotting. In time-lapse movies of cell cultures that showed near-complete depletion of EB1 and EB3 (Fig. 5C), we quantified the time taken for two key events: (i) alignment of last chromosome on metaphase plate (from Nuclear Envelope Break-Down (NEBD) to completion of chromosome congression) and (ii) initiation of chromatid separation (from NEBD to initiation of anaphase onset). Our time-lapse analysis showed a pronounced anaphase onset delay in the vast majority of EB1 and EB3 co-depleted cells but not control-depleted cells. Importantly, comparing the rate and plateau in our timing graphs showed that the delay in anaphase onset was more pronounced compared to the delay in completing chromosome congression (compare Fig. 5D,E). This shows the importance of EB function in timing the onset of anaphase.

The pronounced anaphase delay in EB1 and EB3 co-depleted cells, compared to the noticeable but subtle delay in congression, further supports our model that excessive SKAP-EB interaction induced loss of EB function may be responsible for delaying anaphase without severely disrupting chromosome congression. Thus, the disruption of SKAP-EB interaction and the co-depletion of EB1 and EB3 result in similar mitotic outcomes. This correlative evidence further illustrates the general importance of EB and SXIP-motif mediated plus-end regulation in ensuring the normal timing of anaphase onset.

**DISCUSSION**

To understand how microtubule-ends perform several spatially and temporally distinct tasks, we set out to obtain a comprehensive map of EB1 interactors from two distinct phases of mitosis. This proteome-wide study has unraveled several mitosis phase-specific interactors of EB1 and in addition revealed the Astrin/SKAP complex as a mitotic phase determined interactor of EB1. Investigating the significance of mitotic phase determined SKAP-EB interaction, revealed four unrecognised roles for the SKAP-EB interaction mediating SXIP motif. The SXIP motif controls (i) SKAP’s interaction with EB3 (ii) SKAP’s role in regulating MT growth, (iii) SKAP’s localisation onto spindle microtubules and (iv) SKAP’s role in controlling anaphase onset times. Thus regulated interaction of microtubule plus-end complexes may represent a key rate-limiting step in determining anaphase onset, independent of chromosome congression, and in turn defining mitotic outcome.

Mitotic phase-specific changes in phosphorylation of various microtubule-associated proteins are known (Pagliuca et al., 2011) but it is unclear how these phosphorylations change microtubule-end composition. Phosphorylation of the interphase plus-end binding proteins SLAIN2 (van der Vaart et al., 2011) and CLASP2 (Kumar et al., 2012) that directly interact with EB1, is known to dislodge them from plus-ends during mitosis. Expanding this knowledge, our effort provides a comprehensive list of EB1 interactions that change through mitotic phases. Interaction between CLASP2 and EB1 is likely to be spatially regulated because although CLASP2 is excluded from mitotic plus-ends (Kumar et al., 2012), CLASP2 is recruited to kinetochore-microtubule interface (Pereira et al., 2006) that is enriched for EB1 (Tirnauer et al., 2002), consistent with our finding of CLASP2 in EB1 immunoprecipitates from mitotic cells. Such refined spatial control over plus-end protein interactions during mitosis is probably achieved through localized kinase and phosphatase activities, a poorly understood area (reviewed in Tamura and Draviam, 2012).

Our mitotic-phase enrichment strategy has allowed high-throughput immunoprecipitation studies to reliably capture mitotic-phase associated changes in plus-end complexes. Previous SILAC-based quantitative *in vitro* approaches could distinguish microtubule-binding proteins from interphase *versus* mitotic cells (Syred et al., 2013). Thus, a SILAC-based methodology, in combination with our mitotic-phase enrichment strategy, should be a viable future option.

The outer kinetochore proteins, HEC1Ndc80 and KNL1SPC105 can directly contact the microtubule wall (Cheeseman et al., 2006; Wei et al., 2007). In addition, kinetochore bound motors and microtubule associated proteins can serve as additional molecular bridges between the microtubule and kinetochore (Hsu and Toda, 2011; Jeyaprakash et al., 2012; Maio et al., 2003; Maure et al., 2011; Schmidt et al., 2012; Steuer et al., 1990; Wang et al., 2012; Wood et al., 1997). However, it is unclear how protein-protein interactions at the kinetochore-microtubule interface accommodate structural changes of the growing and shrinking phases of microtubule ends. Our finding that the SXIP-motif of SKAP is dispensable for its recruitment to kinetochores demonstrates that the kinetochore bound pool of SKAP does not require EB1 or EB3 interaction. This is consistent with the recruitment of SKAP to anaphase kinetochores (Dunsch et al., 2011; Fang et al., 2009; Schmidt et al., 2010) that are predominantly tethered to depolymerizing microtubule-ends lacking EB proteins. Thus, our molecular evidence for two pools of SKAP – one that requires regulated SXIP-motif interaction for proper microtubule growth and the other that associates with kinetochore in a SXIP-motif independent manner – reveals the Astrin-SKAP complex as a unique class of outer kinetochore bound microtubule associated protein that arrives at the kinetochores following microtubule-end association (Shrestha and Draviam, 2013) and remains at the kinetochore-microtubule interface regardless of structural changes associated with the presence or absence of microtubule growth associated EB proteins.

Despite SKAP’s ability to directly interact with microtubules *in vitro*, independent of EB1 (Dunsch et al., 2011; Wang et al., 2012), we find that SKAP-EB interaction is essential for SKAP’s localization onto spindle microtubules *in vivo*. SKAP-EB interaction must be finely regulated since we find that excess of SKAP-EB interaction disrupts mitotic timing and microtubule growth rates. We propose that SKAP-EB interaction in prometaphase must be regulated so that EB’s interaction with other EB partners can remain unperturbed (Fig. 5F). In support of this model, the defects in microtubule growth and anaphase onset...
associated with SKAP overexpression are absent in cells overexpressing SKAP(NN) mutant (Fig. 5F) but are present in cells co-depleted of EB1 and EB3 proteins (Ferreira et al., 2013; this study).

Although, our MS studies could not recover any peptide bearing the SXIP motif of SKAP for confirming the phosphorylation status around SKAP’s SXIP motif, we propose that SKAP-EB interaction may be temporally controlled by a mitotic phase-specific kinase or phosphatase for 3 reasons: (i) The proteomic data presented here show the interaction between the Astrin-SKAP complex and EB1 to be mitotic phase dependent. (ii) Our protein-protein interaction studies show that phospho-modulation of an Aurora-B consensus site near the SXIP motif is sufficient for negatively regulating SKAP-EB interaction. (iii) In S. cerevisiae, phosphorylation near the SXIP-motif (Zimiak et al., 2009) is reported to control EB1/Bim1 interaction with AuroraB/Ipl1 in anaphase.

Throughout this study, we recurrently found evidence for spatially and temporally regulated interaction among plus-end proteins during mitosis: in our proteomic studies of mitotic phase-specific EB1 interactors, in our localisation studies of SKAP-EB interaction dependent SKAP enrichment on spindle microtubules, and in our time-lapse studies of SKAP-EB interaction dependent changes in microtubule growth and anaphase onset. Thus, determining mitotic phase-specific EB interactions is a crucial step towards our understanding of how microtubule plus-end complexes execute spatially and temporally distinct mitotic events.

**MATERIALS AND METHODS**

**Cell culture and synchronization**

UTA6 and HeLa cells were cultured in Dulbecco’s modified Eagle’s medium supplemented with 10% FCS and antibiotics, penicillin and streptomycin. Cells were plated onto plastic dishes for large-scale cell culture and glass-bottomed dishes (LabTek) or 13 mm round coverslips for imaging. For inhibition studies, cells were treated with 10 μM Na3VO4, 0.05% (w/v) Triton, 1× Protease inhibitor cocktail (Roche) in PBS (PAA), rotating the tube at 4°C for 3 min, placing tube on magnetic stand and discarding the supernatant. Subsequently the immunoprecipitated protein complexes were eluted with anti-Flag peptide containing elution buffer (50 μl; anti-3×Flag peptide (1 mg/ml) (Invitrogen F4299), 3 mM NaF, 1 mM Na2VO4, 0.05% (w/v) Triton, 1× Protease inhibitor cocktail) by vortexing for 20 min at 4°C and then the elutant was transferred into a new tube. This elution step was repeated 5 times and the elutant and beads were stored at −20°C. To test the efficiency 10% of elutant was mixed with 4×SDS buffer and loaded onto a gel for immunoblotting. The remaining elutant was precipitated with acetone (1:10 elutant:acetone) for 20 min at −20°C and centrifuged (6000 rpm, 10 min), supernatant was removed and pellet dried at room temperature for 10 min. The dry pellets were processed for mass spectrometry analysis at the Institute of Biochemistry and Biophysics in the Polish Academy of Science. For one of the repeats, the mass spectrometry analysis was performed at the Cambridge Centre for Proteomics, University of Cambridge.

**Yeast two hybrid (Y2H) analysis**

Human cDNA fragments encoding for EB1 (NM_012325), EB3 (NM_012326), SKAP (NM_033286.2), Astrin (NM006461.3) were subcloned into pGBT9 and pGAD424 (Clontech). SKAP point mutants (113LP to NN and RAT90 into either RAA or RAE) were created by PCR mutagenesis and confirmed by DNA sequencing. Yeast two-hybrid protocols were based on the Matchmaker 3 yeast two-hybrid system (Clontech).

**Yeast strains and plasmids used in this study**

Yeast strains (AH109 and Y187) and plasmid vectors (pGAD242 and pGBT9) were kindly provided by V Bolanos Garcia (Blundell group, University of Cambridge). N-terminally tagged Flag-EB1 or Flag-Nuf2 were generated using the Tetracycline-repressible vector pTRE-Tight-BI-AcGFP1 (Clontech). N-terminally tagged GFP-SKAP was generated using the Tetracycline-inducible FRT/TO system (Life Technologies).

**Live-cell studies (time-lapse imaging)**

Cells were transfected with siRNA oligos or plasmid vectors for 48 or 24 h, respectively, prior to imaging and transferred to Leibovitz’s L15 medium (Invitrogen) for imaging at 37°C. For live-cell imaging movies of GFP-SKAP(WT) or GFP-SKAP(NN) expressing cells, exposures of 0.02-0.05 s was used for acquiring three Z-planes, 3 μm apart, once every 3 min for 6 to 8 h, with a 40 times NA 0.75 objective. For live-cell imaging movies of EB3-mKate comets, exposures of less than 0.02 s was used for acquiring at least 10 Z-planes, 0.1 μm apart, in continuous acquisition mode, for 5 min, with a 100× NA 1.4 objective. All live-cell imaging studies were performed on an Applied Precision DeltaVision Core microscope equipped with a Cascade2 camera under EM mode.

siRNA transfection

The following published siRNA oligos were used: EB1 oligo (5′-UUGCCUUGAAGAGUUGAA dT.dT-3′; Dharmacon) (Draviam et al., 2006) and EB3 oligo (5′-CAUGAGACUGAUAGCAGAUAUU-3′; Invitrogen) (Ban et al., 2009). siRNA transfection were carried out twice, 48 h and 72 h prior to imaging, using Oligofectamine (Life Technologies) according to manufacturer’s instruction.

**Immunofluorescence and immunoblotting**

For immunofluorescence, antibodies against Tubulin (Abcam; ab6160), Flag (Sigma; F7425), Tubulin (Sigma; T4026), GFP (Roche; 1181446001), SKAP (Atlas; HPA042027), and CREST anti-sera (Europa; FZ90C-CS1058) were used. Images of immunostained cells were acquired using 100 times NA 1.4 objective on a DeltaVision Core microscope equipped with CoolSnap HQ Camera (Photometrics). For immunoblotting, antibodies against Tubulin (Sigma-Aldrich; T6557),...
Draviam, V. M., Shapiro, I., Aldridge, B. and Sorger, P. K. (2006). Misorientation and reduced stretching of aligned sister kinetochore promoters promote chromosome missegregation in EB1- or APC-depleted cells. EMBO J. 25, 2814-2827.


